WORLD INTELLECTUAL PROPERTY ORGANIZATION International Bureau



INTERNATIONAL APPLICATION PUBLISHED UNDER THE PATENT COOPERATION TREATY (PCT)

(51) International Patent Classification ⁶ :	·	(11) International Publication Number: WO 96/34873
C07H 15/12, C12P 21/06, A61K 39/00, 38/00, C07K 1/00	A1	(43) International Publication Date: 7 November 1996 (07.11.96)
(21) International Application Number: PCT/Us (22) International Filing Date: 1 May 1996		CH, DE, DK, ES, FI, FR, GB, GR, IE, IT, LU, MC, NL,
(30) Priority Data: 08/431,517 1 May 1995 (01.05.95)	τ	Published With international search report.
(71) Applicant: INCYTE PHARMACEUTICALS, INC. 3174 Porter Drive, Palo Alto, CA 94304 (US).	[US/US): ·
(72) Inventors: SCOTT, Randal, W.; 13140 Sun-Mohr, View, CA 94040 (US). MARRA, Marian, N.; Harte Drive, Redwood City, CA 94061 (US).		
(74) Agent: GREGG, Valeta; Fish & Richardson P.C., 2200 Sand Hill Road, Menlo Park, CA 94025 (U		
,		
(54) Title: RECOMBINANT ENDOTOXIN-NEUTRAL		

(57) Abstract

In general, the invention features a recombinant endotoxin-neutralizing polypeptide (RENP) characterized by (i) an amino acid sequence, (ii) an amino acid sequence and structure that facilitates selective and specific binding to lipopolysaccharide and (iii) once bound to the lipopolysaccharide, provides endotoxin-neutralizing activity. Preferably, the RENP is composed of an amino acid sequence similar to, but not identical to, an amino acid sequence of BPI, LBP, or both. preferably, the RENP contains an LPS-binding domain derived from the amino acid sequence of BPI, LBP or both. Preferably, the RENPs are covalently bound to a molecule which enhances the half-life of the polypeptide. The RENPs of the invention can be used in pharmaceutical compositions for therapeutic and prophylactic regimens, as well as in various in vitro and in vivo diagnostic methods.

FOR THE PURPOSES OF INFORMATION ONLY

Codes used to identify States party to the PCT on the front pages of pamphlets publishing international applications under the PCT.

AM	Armenia	GB	United Kingdom	MW	Malawi
AT	Austria	GE	Georgia	MX	Mexico
ΑU	Australia	GN	Guinea	NE	Niger
BB	Barbados	GR	Greece	NL	Netherlands
BE	Belgium	HU	Hungary	NO	Norway
BF	Burkina Faso	IE	Ireland	NZ	New Zealand
BG	Bulgaria	IT	Italy	PL	Poland
BJ	Benin	JP	Japan	PT	Portugal
BR	Brazil	KE	Kenya	RO	Romania
BY	Belarus	KG	Kyrgystan	RU	Russian Federation
CA	Canada	KP	Democratic People's Republic	SD	Sudan
CF	Central African Republic		of Korea	SE	Sweden
CG	Congo	KR	Republic of Korea	SG	Singapore
CH	Switzerland	KZ	Kazakhstan	SI	Slovenia
CI	Côte d'Ivoire	u	Liechtenstein	SK	Slovakia
CM	Cameroon	LK	Sri Lanka	SN	Senegal
CN	China	LR	Liberia	SZ	Swaziland
CS	Czechoslovakia	LT	Lithuania	TD	Chad
CZ	Czech Republic	LU	Luxembourg	TG	Togo
DE	Germany	LV	Latvia	TJ	Tajikistan
DK	Denmark	MC	Monaco	TT	Trinidad and Tobago
EE	Estonia	MD	Republic of Moldova	UA	Ukraine
ES	Spain	MG	Madagascar	UG	Uganda
FI	Finland	ML	Mali	US	United States of America
FR	France	MN	Mongolia	UZ	Uzbekistan
GA	Gabon	MR	Mauritania	VN	Viet Nam

RECOMBINANT ENDOTOXIN-NEUTRALIZING PROTEINS

Field of the Invention

This invention relates generally to the field of recombinant, endotoxin-neutralizing proteins, particularly to recombinant proteins which bind endotoxin and block endotoxin-mediated activation of biological systems.

10 Background of the Invention

Gram-negative infections are a major cause of morbidity and mortality, especially in hospitalized and immunocompromised patients. [Duma, Am. J. of Med., 78 (Suppl. 6A):154-164 (1985); and Kreger et al., Am. J.

- 15 Med., 68:344-355 (1980)]. Although available antibiotics are generally effective in inhibiting growth of Gram-negative bacteria, they do not neutralize the pathophysiological effects associated with endotoxins. Endotoxin is a heat stable bacterial toxin composed of
- 20 lipopolysaccharides (LPS) released from the outer
 membrane of Gram-negative bacteria upon lysis
 [Shenep et al., J. Infect. Dis., 150(3):380-388 (1984)],
 and is a potent stimulator of the inflammatory response.
 Endotoxemia occurs when endotoxin enters the bloodstream
 25 resulting in a dramatic systemic inflammatory response.

Many detrimental in vivo effects of LPS result from soluble mediators released by inflammatory cells. [Morrison et al., Am. J. Pathol., 93(2):527-617 (1978)]. Monocytes and neutrophils, which ingest and kill microorganisms, play a key role in this process.

Monocytes and neutrophils respond to endotoxin in vivo by releasing soluble proteins with microbicidal,

proteolytic, opsonic, pyrogenic, complement-activating and tissue-damaging ff cts. These factors mediate many of the pathophysiological effects of endotoxin. For example, tumor necrosis factor (TNF), a cytokine released by endotoxin-stimulated monocytes, causes fever, shock, and alterations in glucose metabolism and is a potent stimulator of neutrophils. Other cytokines such as IL-1, IL-6, and IL-8 also mediate many of the pathophysiologic effects of LPS, as well as other pathways involving endothelial cell activation by tissue factor, kininogen, nitric oxide and complement.

Endotoxin-associated disorders result from extra-gastrointestinal exposure to LPS, e.g. administration of LPS-contaminated fluids, or 15 Gram-negative infections. Endotoxin-associated disorders can also result when the natural epithelial barrier is injured and the normal Gram-negative flora breach this barrier. For example, endotoxin-associated disorders can occur (a) when there is ischemia of the gastrointestinal 20 tract (e.g., following hemorrhagic shock or during certain surgical procedures), or (b) when systemic or local inflammation causes increased permeability of the gut to endotoxin or Gram-negative organisms. presence of endotoxin and the resulting inflammatory 25 response may result, for example, in endotoxemia, systemic inflammatory response syndrome (SIRS), sepsis syndrome, septic shock, disseminated intravascular coagulation (DIC), adult respiratory distress syndrome (ARDS), cardiac dysfunction, organ failure, liver failure 30 (hepatobiliary dysfunction), brain failure (CNS dysfunction), renal failure, multi-organ failure and shock.

Examples of diseases which can be associated with Gram-negative bacterial infections or endotoxemia include bacterial meningitis, neonatal sepsis, cystic fibrosis,

- 3 -

inflammatory bowel disease and liver cirrhosis,
Gram-negative pneumonia, Gram-negative abdominal abscess,
hemorrhagic shock and disseminated intravascular
coagulation. Subjects who are leukopenic or neutropenic,
including subjects treated with chemotherapy or
immunocompromised subjects (for example with AIDS), are
particularly susceptible to bacterial infection and the
subsequent effects of endotoxin.

Several therapeutic compounds have been developed 10 to inhibit the toxic effects of endotoxin, including antibacterial LPS-binding agents and anti-LPS antibodies, although each has met with limitations. For example, Polymyxin B (PMB) is a basic polypeptide antibiotic which binds to Lipid A, the most toxic and biologically active 15 component of endotoxin. PMB inhibits endotoxin-mediated activation of neutrophil granule release in vitro and is a potential therapeutic agent for Gram-negative infections. However, because of its systemic toxicity, this antibiotic has limited therapeutic use, and is 20 generally used topically. Combination therapy using antibiotics and high doses of methylprednisolone sodium succinate (MPSS) showed more promise as this regimen prevented death in an experimental animal model of Gram-negative sepsis. However, a clinical study using 25 MPSS with antibiotics in treatment of patients having clinical signs of systemic sepsis showed that mortality rates were not significantly different between the treatment and placebo groups [Bone et al., N. Engl. J. Med. 317:653 (1987)].

Antibodies that bind endotoxin have been used in the treatment of endotoxemia. For example, hyperimmune human antisera against E. coli J5 reduced mortality by 50% in patients with Gram-negative bacteremia and shock [Ziegler et al., N. Engl. J. Med. 307:1225 (1982)].

35 However, attempts to treat Gram-negative sepsis by

administration of anti-LPS monoclonal antibodies met with little or no success [Ziegler et al., N. Engl. J. Med. 324:429 (1991); Greenman et al., JAMA 266:1097 (1991); Baumgartner et al., N. Engl. J. Med. 325:279 (1991)].

5

Another approach to treating endotoxemia involves the use of cytokine blockers, such as IL-1 receptor antagonists and anti-TNF antibodies, as well as the soluble forms of the IL-1 and TNF receptors. However. any given cytokine blocker blocks only the cytokine for 10 which it is specific, and fails to prevent the action of other cytokines. Furthermore, blocking cytokines may have other deleterious effects.

Two soluble endotoxin-binding proteins, lipopolysaccharide binding protein (LBP) and 15 bactericidal/permeability-increasing (BPI), play opposing roles in vivo in the physiological response to endotoxin. LBP is a soluble LPS receptor found in serum which binds LPS with high affinity via interaction with the Lipid A moiety [Tobias et al. (1986) J. Exp. Med. 164:777-793; 20 Tobias et al. (1989) J. Biol. Chem. 264:10867-10871]. LBP-LPS complexes stimulate monocyte activation through interaction with the CD14 receptor on the surface of monocytes, resulting in production of cytokines such as TNF and IL-1 [Wright et al. (1989) J. Exp. Med. 25 170:1231-1241; Wright et al. (1990) Science 249:1431]. Thus, LBP acts as a transfer protein in LPS-mediated stimulation of cytokine release. Moreover, LBP increases

In direct contrast to LBP, BPI binds and neutralizes endotoxin, preventing inflammatory cell activation. BPI, also known as CAP57 and BP [Shafer et al., Infect. Immun. 45:29 (1984); 35 Hovde et al., Infect. Immun. 54:142 (1986)] is also

30 than in its absence.

LPS activity in that a lower concentration of LPS is required to stimulate monocytes in the presence of LBP

- 5 -

bactericidal by virtue of its interaction with the
Lipid A moi ty of LPS in th bacterial cell wall. BPI
binds LPS, disrupts LPS structure and the cell wall, and
increases bacterial membrane permeability, resulting in
5 cell death [Weiss et al., J. Biol. Chem, 253:2664-2672
(1978); Weiss et al., Infection and Immunity 38:1149-1153
(1982)]. BPI retains its in vitro bactericidal activity
after protease cleavage, suggesting that BPI fragments
retain activity [Ooi et al., Clinical Research 33(2):567A
10 (1985)]. This observation was confirmed by Ooi et al.,
who showed that an N-terminal 25 kD fragment of BPI
exhibited both the in vitro bactericidal and permeability
increasing activities [Ooi et al., J. Biol. Chem.
262:14891 (1987)].

15 Molecular Structures of BPI and LBP

The genes encoding BPI and LBP have been cloned (Gray et al. (1989) J. Biol. Chem. 264:9505-9509; Schumann et al., Science 249:1429-1431 (1990)]. BPI and LBP are immunologically cross-reactive, contain a 20 hydrophobic leader sequence, and share significant amino acid sequence homology over the entire length of the molecules, with an overall amino acid sequence identity of 44% [Tobias et al., J. Biol. Chem. 263:13479-13481 (1988); Schumann et al. supra]. BPI and LBP each contains three cysteine residues. BPI contains two glycosylation sites; LBP contains five potential glycosylation sites.

BPI is characterized by two distinct domains, an N-terminal domain and a C-terminal domain, which are separated by a proline-rich hinge region. The N-terminal domain of BPI has strong LPS-neutralizing activity, while the C-terminal domain of BPI has modest LPS-neutralizing activity. LBP can also be divided into N- and C-terminal domains, with the C-terminal domain being implicated in

- 6 -

binding of LPS to macrophages and their subsequent activation.

The N- and C-terminal domains of BPI have a striking charge asymmetry that is not shared by LBP. The 5 N-terminal domain of BPI, which is rich in positively charged lysine residues, imparts a predicted pI > 10 to the full-length molecule. In contrast, the C-terminal domain of BPI is only slightly negatively charged. LBP, which is a neutral protein, has no bactericidal activity [Tobias et al., J. Biol. Chem. 263:13479 (1988)]. This suggests that the bactericidal activity of BPI results from its overall cationicity.

Table 1 provides a comparison of BPI and LBP structure and function.

. - /

Table 1
Comparison of BPI and LBP Structure and Function

		BPI	LBP
	SYNTHESIS		
	Site of synthesis	Neutrophil	Liver
5	Blood concentration	1-10 ng/ml	1-10 μg/ml
	STRUCTURE		
	Molecular mass	55 kD	60 kD
	Glycosylation sites	2	5
	Cysteine	3	3
10	EFFECTS ON LPS MEDIATED:		
	neutrophil activation	Inhibits	Stimulates
	monocyte activation	Inhibits	Stimulates
	TNF release	Inhibits	Stimulates
	IL-1 release	Inhibits	Stimulates
15	IL-6 release	Inhibits	Stimulates

^{*} Four cysteines were reported by Schumann et al. [Science 249:1429-1431 (1990)]. Subsequent DNA sequence analysis by the present inventors determined that Schumann's sequence was erroneous and that LBP contains only three cysteine residues (see Figure 1).

Therapeutic intervention to block the inflammatory effects of LPS would ameliorate the morbidity and mortality associated with endotoxemia and septic shock. Unfortunately, although BPI binds LPS with high affinity, it has an extremely short half-life in vivo, thus limiting its use in therapy. Native LBP has a longer half-life but, upon binding of LPS, elicits a brisk monocyte reaction which can facilitate release of deleterious quantities of cytokines.

Early and specific diagnosis of endotoxin-associat d disord rs is essential in the identification of patients who have or who are at risk of developing such disorders.

Precise identification of a site of Gram-negative infection in a patient would assist the clinician in the design and targeting of antibacterial therapy.

5

An ideal anti-endotoxin drug candidate and/or LPS detection reagent would have a longer half-life and 10 effective, high-affinity endotoxin binding/inactivation without monocyte stimulation. There is a clear need in the field for specific diagnostic and therapeutic agents which neutralizes the effects of endotoxin and has an acceptably long half-life in vivo. The present invention 15 addresses these problems.

Summary of the Invention

Recombinant proteins are genetically engineered to bind lipopolysaccharide (LPS) such that the endotoxin is inactivated, thus preventing the endotoxin from inducing the immunological cascade of events associated with endotoxin-related disorders (e.g., activation of monocytes, tumor necrosis factor (TNF) production).

In general, the invention features a recombinant endotoxin-neutralizing polypeptide (RENP) characterized by (i) an amino acid sequence, (ii) a sequence and structure that facilitate specific binding to lipopolysaccharide, (iii) provides endotoxin-neutralizing activity upon LPS binding, and (iv) a half-life that is enhanced relative to the half-life of BPI. Preferably, the RENP is composed of an amino acid sequence similar to, but not identical to, an amino acid sequence of BPI, LBP, or both. Preferably, the RENP contains an LPS-binding domain derived from the amino acid sequence of BPI, LBP, or both. Preferred RENPs are fusion

- 9 -

proteins which bind LPS with the high affinity of BPI, but do not contain the BPI amino acid sequences associated with BPI's short half-life.

Preferably, the RENPs are covalently bound to a

5 molecule which further enhances the half-life of the
polypeptide. For example, the half-life enhancing
molecule can be an immunoglobulin fragment, a half-life
determining portion of LBP or LBP derivative, or
polyethylene glycol. In related aspects, the invention

10 features DNA encoding an RENP of the invention, vectors
and transformed cells containing DNA encoding an RENP, a
method for producing RENPs, and detectably labeled RENPs.

A primary object of the invention is to provide an RENP which binds and inactivates endotoxin, and has a half-life suitable for administration to a patient.

Another object of the invention is to provide a pharmaceutical composition containing a therapeutically effective amount of an RENP for use in treatment of endotoxin-related disorders.

20 Still another object of the invention is to provide endotoxin-neutralizing proteins for use in the detection of LPS. The RENPs can be bound to a label which can be detected or can be bound to a support for use in LPS-detection assays. LPS can be detected in vivo to identify a site of infection in a subject or can be used in an in vitro assay to qualitatively or quantitatively detect LPS in a sample.

Another object of the invention is to provide endotoxin-neutralizing proteins that can be used to 30 produce endotoxin-free solutions and tools for use in, for example, various medical applications.

An advantage of the present invention is that the endotoxin-neutralizing proteins have a half-life in serum which is enhanced relative to the half-life of naturally-occurring LPS-binding proteins, and bind LPS

. - 10 -

without triggering a significant, undesirable immune r sponse.

Another advantage of the invention is that the RENPs can be administered to a patient immediately upon identification of a symptom of an endotoxin-associated disorder.

Another advantage is that the endotoxin-neutralizing proteins can be administered prophylactically to a patient at risk of endotoxic shock or other LPS-mediated condition.

An advantage of the invention is that various RENPs having binding specificity for LPS for detection of LPS either in vivo or in vitro.

Another advantage of the invention is that the 15 RENPs can be attached to a variety of detectable labels.

Yet another advantage of the invention is that the RENPs can be bound to a molecule which can interact with or which can be a portion of a solid support.

These and other objects, advantages and features
of the present invention will become apparent to those
persons skilled in the art upon reading the details of
the vectors, cell lines and methodology as more fully set
forth below.

Brief Description of the Drawings

Figures 1A-1D are a comparison of the amino acid sequences of human LBP as described by Schumann et al. (LBPa) and as used herein (LBPb).

Figure 2 is a schematic diagram showing the various combinations of BPI, LBP, BPI variants, and/or 30 LBP variants which can be used to generate an RENPs of the invention.

Figures 3A-3D show the nucleotide and amino acid sequences of BPI.

Figures 4A-4C show the nucleotide and amino acid sequences of LBP.

Figures 5A-5F are a comparison of the amino acid sequences of BPI and LBP from various species.

5 Figure 6 shows the amino acid sequence of $L_{1-197}B_{200-456}$ (NCY118).

Figure 7 is a graph showing the effects of BPI, LBP, $L_{1-197(143->V)}B_{200-456(N206->D)}$ (NCY103) and $B_{1-199}L_{200-456}$ (NCY104) on biotinylated BPI binding to LPS.

10 Figure 8 is graph showing the effects of BPI, LBP, $L_{1-197(143-v)}B_{200-456(N206-v)}$ (NCY103), $B_{1-199}L_{200-456}$ (NCY104), or $B_{(8351-v)}$ (NCY105) protein on LPS activity in the chromogenic LAL assay.

Figure 9 is a graph showing FITC-LPS binding to monocytes in the presence of BPI or $L_{1-197(143->V)}^{B}_{200-456(N206->D)}$ (NCY103).

20

Figure 10 is a graph showing the effects of BPI, LBP, $L_{1-197(143->V)}B_{200-456(N206->D)}$ (NCY103) or $B_{1-199}L_{200-456}$ (NCY104), on TNF release by LPS in whole blood.

Figure 11 is a graph showing clearance of BPI, LBP, $L_{1-197(143-v)}^{B_{200-456(N206-v)}}$ (NCY103) or $B_{1-199}^{L_{200-456}}$ (NCY104) from mouse serum after intravenous injection.

Figure 12 is a graph comparing the efficacy of BPI and $L_{1-197(143->V)}B_{200-456(N206->D)}$ (NCY103) in the protection 25 to endotoxin challenge.

Figures 13A-13C are graphs showing the effects of BPI, $L_{1-197}(143-v)^B_{200-456}(N206-v)$ (NCY103), $L_{1-197}^B_{200-456}$ (NCY118), $L_{1-198}^B_{201-456}$ Fc (NCY144), $L_{1-59}^B_{60-456}$ (NCY114), $L_{1-134}^B_{135-456}$ (NCY115), $L_{1-359}^B_{360-456}$ (NCY117), and B_{CAT9} 30 (NCY139) on biotinylated BPI binding to LPS.

Figures 14A-14B are graphs showing the effects of BPI, LBP, $L_{1-197(143->V)}B_{200-456(N206->D)}$ (NCY103) and $B_{1-199}L_{200-456}$ (NCY104) on FITC-labeled LPS binding to human peripheral blood monocytes in the presence of 10%

autologous serum (14A) and in the absence of serum and presence of 0.5% human serum albumin (14B).

Figure 15 is a graph comparing the effects of LBP vs. $L_{1-197(143-v)}B_{200-456(N206-v)}$ (NCY103), $B_{1-199}L_{200-456}$ (NCY104), $L_{1-359}B_{360-456}$ (NCY117) and PLL (poly-L-lysine) on the stimulation of TNF α release by phorbol ester-induced THP-1 cells in the absence of serum.

Figure 16 is a graph showing the effects of various recombinant-endotoxin neutralizing proteins upon 10 LPS-mediated TNF production in THP-1 cells cultured without serum.

Figures 17A-17H are graphs showing the clearance of: BPI, LBP, $L_{1-197}(143-v)^B_{200-456}(N206-v)$ (NCY103), $B_{1-199}L_{200-456}$ (NCY104), and $L_{1-197}B_{200-456}$ (NCY118) (17A); 15 BPI, $L_{1-59}B_{60-456}$ (NCY114), $L_{1-134}B_{135-456}$ (NCY115), and B_{CAT9} (NCY139) (17B); BPI, LBP, $L_{1-359}B_{360-456}$ (NCY117) and $L_{1-197}B_{200-456}$ (NCY118) (17C); and BPI, LBP and $L_{(1-198)}B_{(201-456)}$ FC (NCY144) (assayed for both Fc and BPI) in CD-1 mice (17D); LBP, $L_{1-275}B_{278-456}$ (NCY116), 20 $L_{1-359}B_{360-456}$ (NCY117), $L_{1-197}B_{200-456}$ (NCY118) (17E); LBP, $L_{1-197}(143-v)^B_{200-456}(NCY117)$, $L_{1-197}B_{200-456}$ (NCY118) (17E); LBP, $L_{1-197}(143-v)^B_{200-456}(NC206-v)$ (NCY103), $L_{1-134}B_{135-456}$ (NCY115), $L_{(1-198)}B_{(202-275)}L_{(274-466)}$ (NCY135), and

(NCY141), L_{CAT9} (NCY142), L_{CAT15} (NCY143) and BPI (17G); 25 and BPI, $L_{1-134}B_{135-456}$ (NCY115), and $L_{1-59}B_{60-456}$ (NCY114) (17H).

 $L_{(1-134)}B_{(136-275)}L_{(274-456)}$ (NCY134) (17F); LBP (NCY102), L_{CAT6}

Figure 18 is Western blot of BPI and $L_{1-197}B_{200-456}$ (NCY118) produced in *Pichia pastoris*.

Figure 19 is a graph showing the effects of BPI and $L_{1-197(143->V)}B_{200-456(N206->D)}$ (NCY103) on endotoxin activation of monocytes.

Figure 20 is a graph showing the protective effects of $L_{1-197}B_{200-456}$ (NCY118) to endotoxin challenge in mice.

- 13 -

Description of the Preferred Embodiments

endotoxin-neutralizing proteins, methods for providing therapy to a patient suffering from an endotoxin-related 5 disorder, and compositions and method for diagnosis of a condition associated with LPS are described, it is to be understood that this invention is not limited to the particular methodology, protocols, cell lines, vectors and reagents described as such may, of course, vary. It is also to be understood that the terminology used herein is for the purpose of describing particular embodiments only, and is not intended to limit the scope of the present invention which will be limited only by the appended claims.

It must be noted that as used herein and in the appended claims, the singular forms "a", "and", and "the" include plural referents unless the context clearly dictates otherwise. Thus, for example, reference to "a recombinant endotoxin-neutralizing protein" includes a plurality of such proteins and reference to "the DNA encoding the recombinant endotoxin-neutralizing protein" includes reference to one or more transformation vectors and equivalents thereof known to those skilled in the art, and so forth.

Unless defined otherwise, all technical and scientific terms used herein have the same meaning as commonly understood to one of ordinary skill in the art to which this invention belongs. Although any methods, devices and materials similar or equivalent to those described herein can be used in the practice or testing of the invention, the preferred methods, devices and materials are now described.

All publications mentioned herein are incorporated herein by reference for the purpose of describing and 35 disclosing the cell lines, vectors, and methodologies

which are described in the publications which might be used in connection with the presently d scribed invention.

Definitions

By "lipopolysaccharide" or "LPS" is meant a compound composed of a heteropolysaccharide (which contains somatic O antigen) covalently bound to a phospholipid moiety (lipid A). LPS is a major component of the cell wall of Gram-negative bacteria.

By "endotoxin" is meant a heat-stable toxin 10 associated with the outer membranes of certain Gram-negative bacteria, including the enterobacteria, brucellae, neisseriae, and vibrios. Endotoxin, normally released upon disruption of the bacterial cells, is 15 composed of lipopolysaccharide molecules (LPS) and any associated proteins. The phospholipid moiety of LPS, lipid A, is associated with LPS toxicity. When injected in large quantities endotoxin produces hemorrhagic shock and severe diarrhea; smaller amounts cause fever, altered 20 resistance to bacterial infection, leukopenia followed by leukocytosis, and numerous other biologic effects. Endotoxin is a type of "bacterial pyrogen," which is any fever-raising bacterial product. The terms "endotoxin," "LPS," and "lipopolysaccharide" as used herein are 25 essentially synonymous.

By "recombinant endotoxin-neutralizing polypeptide", "RENP" or "recombinant LPS-neutralizing polypeptide" is meant a protein which has been genetically engineered and contains an LPS-binding 30 domain. Preferably, such recombinant LPS-binding proteins bind endotoxin, have a relatively long half-life in serum (e.g., compared to bactericidal/permeability increasing (BPI) protein), and elicit no or relatively little of the undesirable inflammatory side effects

associated with endotoxin and/or binding of LPS to particular naturally occurring endotoxin-binding proteins (e.g., lipopolysaccharide binding (LBP) protein).

"RENPs" of the invention do not occur naturally and are distinct from those endotoxin-binding proteins that do occur in nature, specifically BPI and LBP.

By "LPS-binding domain" is meant an amino acid sequence which confers specific and selective LPS binding upon a polypeptide.

By "high affinity LPS binding" is meant an LPS binding affinity greater than the LPS binding affinity of LBP, preferably about the same or greater than the LPS binding affinity of BPI.

By "endotoxin-neutralizing activity" is meant a

15 biological activity associated with inhibition of the
toxic effects of lipopolysaccharide, e.g., by binding LPS
and preventing interaction of LPS with proteins and/or
receptors which mediate an undesirable immunological
response associated with endotoxin in a mammalian host.

By "recombinant" or "genetically engineered" is meant a DNA sequence, or a polypeptide encoded thereby, generated using nucleic acid manipulation techniques (e.g., cloning, PCR, and/or fusion protein techniques). "Recombinant" or "genetically engineered" DNA, and thus the proteins encoded by such DNAs, do not occur in nature.

By "half-life" is meant the time required for a living tissue, organ, or organism to eliminate one-half of a substance introduced into it.

30 By "molecule which enhances the half-life" or "half-life enhancing molecule" is meant chemical moiety (e.g., bound via a chemical modification) which enhances the biological half-life of a polypeptide with which it is associated relative to the biological half-life of the parent polypeptide. Chemical moieties include an amino

- 16 -

acid sequence or prot in. For example, where a polyethylene glycol (PEG) moi ty is covalently bound to a protein so as to increase the half-life of the protein relative to the un-PEGylated parent protein, the PEG 5 moiety is the "molecule which enhances the half-life" of the protein.

By "half-life determining portion" of a polypeptide is meant an amino acid sequence which is associated with the biological half-life of the 10 polypeptide.

By "bactericidal/permeability increasing protein" or "BPI" is meant a naturally occurring or recombinantly expressed protein having the DNA and amino acid sequences shown in Figures 3A-3D.

By "lipopolysaccharide binding protein" or "LBP" is meant a naturally occurring or recombinantly expressed protein having the DNA and amino acid sequences shown in Figures 1A-1D and Figures 4A-4C.

By "BPI variant" is meant a protein having an amino acid sequence similar to, but not identical to, the amino acid sequence of BPI. "BPI variants" (a) bind LPS, (b) competitively bind LPS in the presence of BPI or LBP, and (c) inhibit the LPS-mediated production of TNFα by human monocytes. In general, "BPI variants" contain the amino acid sequence of BPI but with at least one of: 1) an amino acid substitution; 2) an amino acid deletion; or 3) an amino acid addition, relative to the BPI amino acid sequence.

By "LBP variant" is meant a protein having an amino acid sequence similar to, but not identical to, the amino acid sequence of LBP. "LBP variants" (a) bind LPS, (b) competitively bind LPS in the presence of BPI or LBP, and (c) inhibits production of TNFα by human monocytes. In general, "LBP variants" contain the amino acid sequence of LBP but with at least one of: 1) an amino

- 17 -

acid substitution; 2) an amino acid deletion; or 3) an amino acid addition, relativ to the LPB amino acid sequence.

By "detectable label" is meant any molecule

5 recognized in the art as a means for identifying and/or
detecting a protein to which the detectable label is
bound. Exemplary "detectable labels" include
radionucleotides, fluorescent moieties, biotin, and
antigenic molecules (e.g., a polypeptide which is

10 specifically bound by an anti-polypeptide antibody).
"Detectable labels" include a portion of a chimeric
protein where a portion of the chimeric protein can be
detected by, for example, binding of a detectably labeled
antibody or other detectably labeled molecule which

15 specifically binds the chimeric protein portion.

By "support" is meant a surface to which LPS or an RENP of the invention can be bound and immobilized. Exemplary supports include various biological polymers and non-biological polymers.

20 By "condition associated with endotoxin". "endotoxin associated disorder", or "endotoxin-related disorder" is meant any condition associated with extra-gastrointestinal (e.g., mucosal, blood-borne, closed space) lipopolysaccharide, e.g., a condition 25 associated with bacteremia or introduction of lipopolysaccharide into the blood stream or onto an extra-gastrointestinal mucosal surface (e.g., the lung). Such disorders include, but are not limited to. endotoxin-related shock, endotoxin-related disseminated 30 intravascular coagulation, endotoxin-related anemia. endotoxin-related thrombocytopenia, endotoxin-related adult respiratory distress syndrome, endotoxin-related renal failure, endotoxin-related liver disease or hepatitis, systemic immune response syndrome (SIRS) 35 resulting from Gram-negative infection, Gram-negative

neonatal sepsis, Gram-negative meningitis, Gram-negative pneum nia, neutropenia and/ r leucopenia resulting from Gram-negative infection, hemodynamic shock and endotoxin-related pyresis.

By "transformation" is meant a permanent genetic change induced in a cell following incorporation of new DNA (i.e., DNA exogenous to the cell). Where the cell is a mammalian cell, the permanent genetic change is generally achieved by introduction of the DNA into the 10 genome of the cell.

5

By "transformed cell" is meant a cell into which (or into an ancestor of which) has been introduced, by means of recombinant DNA techniques, a DNA molecule encoding a protein of interest.

By "promoter" is meant a minimal DNA sequence 15 sufficient to direct transcription. "Promoter" is also meant to encompass those promoter elements sufficient for promoter-dependent gene expression controllable for cell-type specific, tissue-specific or inducible by 20 external signals or agents; such elements may be located in the 5' or 3' regions of the native gene.

By "operably linked" is meant that a DNA sequence and a regulatory sequence(s) are connected in such a way as to permit gene expression when the appropriate 25 molecules (e.g., transcriptional activator proteins) are bound to the regulatory sequence(s).

By "operatively inserted" is meant that the DNA of interest introduced into the cell is positioned adjacent a DNA sequence which directs transcription and 30 translation of the introduced DNA (i.e., facilitates the production of, e.g., a polypeptide encoded by a DNA of interest).

By "mammalian subject" or "mammalian patient" is meant any mammal for which the therapy of the invention

- 19 -

is desired, including human, bovine, equine, canine, and felin subjects.

The invention will now be described in further detail.

5 Nomenclature used to describe RENPs

In order to facilitate the discussion and description of the RENPs of the invention, each RENP is designated a specific formula to briefly describe the amino acid sequence of the protein, as well as the origin 10 of specific portions of the protein. The portion of BPI in the recombinant protein is designated with the letter B, followed by an amino acid sequence numbering assignment corresponding to that shown in Figures 5A-5F for human BPI, wherein the mature N-terminus is 15 designated as residue 1. The portion of LBP in certain LBP variants and chimeras is designated by the letter L, followed by an amino acid sequence numbering assignment corresponding to that shown in Figures 1A-1D for human LBP, wherein the mature N-terminus is designated as 20 residue 1. To avoid confusion between the erroneous LBP amino acid sequence published by Schumann et al., supra (designated LBP-a) and the correct LBP amino acid sequence used in the RENPs of the invention (designated LBP-b) and presented in Figures 1A-1D. The differences 25 between the DNA and amino acid sequences for "LBP-a" and

As an example of RENP nomenclature, $L_{1-197}B_{200-456}$ (NCY118) contains amino acid residues 1-199 of LBP fused at the C-terminus of the LBP portion to the N-terminus of amino acid residues 200-456 of BPI. $L_{1-197}B_{200-456}$, shown in Figure 6 has the N-terminal domain of LBP (having a putative endotoxin-binding domain) fused to the C-terminal domain of BPI (having a putative LPS-clearing domain).

"LBP-b" are presented in Table 2A below.

- 20 -

In this application, single amino acid residue substituti ns are noted in parentheses, wherein the original amino acid residue is indicated (using the standard one letter code for amino acids), followed by the substitute amino acid residue. For example, the BPI variant having an alanine residue substituted for the original serine residue at position 351 (which substitution removes a glycosylation signal) is designated BPI_(8351->A). In another example, in

10 B_(DS200->DP), a proline residue is substituted for the serine residue at position 200. In this latter example, the amino acid substitution produces a formic acid-cleavable site.

As another example, the RENP LBP-BPI chimera

15 NCY103 is designated L_{1-198(I43->V)}B_{201-456(D206->N)}. In the recombinant protein, the original isoleucine residue at position 43 of the LBP portion is substituted with a valine residue, and the original asparagine residue at position 206 of the BPI portion is substituted with an 20 aspartate residue. The C-terminus of the LBP amino acid sequence 1-198 having isoleucine substituted at position 43 is covalently bound to the N-terminus of the BPI amino acid sequence 201-456 having valine substituted at position 206.

The amino acid substitutions may be substitutions wherein an original amino acid residue at a given position is substituted with the residue at the corresponding position in a different protein. BPI(In->Y) is an example of such a substitution, wherein amino acid residue X at position n in BPI is substituted with residue Y which is found at position n in LBP (or rabbit or bovine LBP). "X" and "Y" denote amino acid positions in a primary amino acid sequence. "Y" as used in this context is not to be confused with the symbol "Y"

35 denoting the amino acid residue tyrosine. LBP(In->Y) is

- 21 -

another example of such a substitution, wherein amino acid residue X at position n in LBP is substituted with residue Y which is found at position n in BPI (or rabbit or bovine BPI).

Amino acid residue insertion changes are noted in parentheses, by indicating the amino acid residue after which the insertion occurs, followed by the amino acid residue after which the insertion occurs together with the inserted residue or residues. For example,

10 B_{(D200papain}) indicates that an amino acid sequence for cleavage of the BPI variant by papain is inserted after the aspartic acid at residue position 200.

TABLE 2A

Individual Sequence Differences Between
Schumann et al. and LBP as Used Herein

	NUCLEIC ACID		PROTEIN		
	Alpha	Beta	Alpha	Beta	
	A ₄₂	C ₄₂	G ₁₂₉ YCL ₁₃₂	V ₁₂₉ TAS ₁₃₂	
	C318	T318	S ₁₄₉	F ₁₄₉	
0	G488	(np)	A ₂₄₁	V241MSLP245	
	(np)	C499	L411	F411	
	T ₅₄₆	C ₅₄₆			
	C ₅₄₈	T ₅₄₈			
	(np)	T ₈₂₄ CATGAGCCTTC ₈₃			
5	C ₁₃₃₃	T ₁₃₃₃			

(np) = not present in the sequence

15

Table 2B describes some exemplary general classes of RENPs of the invention. In the formulas in Table 2B, n represents an amino acid residue position in the mature 30 sequence of BPI or LBP, x represents an amino acid r sidue in a position which is C-terminal to n in the

sequence of BPI or LBP, and y represents an amino acid residue in a position which is C-terminal to x in the sequence of BPI or LBP. The symbols n, x and y denote the amino acid residue positions as they occur in the mature sequence of the native protein, and not necessarily the positions as they occur in the variant protein.

Table 2B

10

Examples of RENPs

```
BPI variant (N-terminal frag.)
                                                       B_{1-n}
      LBP variant (N-terminal frag.)
                                                       L<sub>1-n</sub>
      BPI variant (C-terminal frag.)
                                                       B<sub>n-456</sub>
      LBP variant (C-terminal frag.)
                                                       Ln-456
·15 BPI variant (internal frag.)
                                                       B_{n-x}
      LBP variant (internal frag.)
      LBP-BPI chimera
                                                       \frac{L_{n-x}B(x+1)-y}{B_{n-x}L(x+1)-y}
      BPI-LBP chimera
                                                      Bn-xL(x+1)-y
Ln-xB(x+1)-456
Bn-xL(x+1)-456
L1-nB(n+1)-x
B1-nL(n+1)-x
L1-nB(n+1)-456
B1-nL(n+1)-456
L1-nB(n+1)-456
B1-nL(n+1)-xL(x+1)-456
B1-nL(n+1)-xL(x+1)-456
      LBP-BPI chimera
 20 BPI-LBP chimera
      LBP-BPI chimera
      BPI-LBP chimera
      LBP-BPI chimera
      BPI-LBP chimera
 25 LBP-BPI-LBP chimera
      BPI-LBP-BPI chimera
                                                       B_{1-n}L_{(n+1)-x}B_{(x+1)-456}
```

All of the constructs in Table 2B can also contain additional molecules which confer an enhanced half-life 30 upon the RENP (e.g., the RENP can be covalently bound to a polyethylene glycol moiety, or a portion of an immunoglobulin protein or other amino acid sequence which confers a half-life increased relative to the unmodified protein). The general scheme for generation of RENPs is outlined in Figure 2.

.- 23 -

Production of RENPs

The RENPs of th invention minimally have characteristics associated with (i) specific and high affinity binding to lipopolysaccharide and 5 (ii) endotoxin-neutralizing activity. In general, the amino acid sequence of RENPs is based upon an amino acid sequence of BPI, LBP, or both. However, the amino acid sequences of the RENPs are distinct from that of BPI and LBP, i.e. the RENPs contain amino acid substitutions. 10 deletions, and/or additions relative to the amino acid sequence of BPI or LBP. Thus, the RENPs of the invention contain: 1) amino acid sequences of a naturally-occurring LPS-binding protein (i.e., LBP and/or BPI); and/or 2) amino acid sequences which do not occur 15 within a single naturally-occurring LPS-binding protein (i.e., LBP or BPI). RENPs can thus be similar to, but not identical to, LBP or BPI. For example, the RENPs can be fragments of BPI and/or LBP, as the amino acid sequences of such RENPs are similar to, but not identical 20 to, naturally occuring BPI or LBP. Moreover, the RENPs of the invention generally have biological properties distinct from and advantageous to either BPI or LBP. RENPs of the invention include BPI variants, LBP variants, and chimeric proteins composed of amino acid 25 sequences derived from BPI, LBP, BPI variants, and/or LBP variants.

For example, RENPs can contain an amino acid sequence of BPI, where the BPI amino acid sequence 1) has been altered at a site of glycosylation (e.g., insertion or deletion of a glycosylation site); 2) contains a neutral or anionic amino acid substituted at a cationic residue of the BPI amino acid sequence (cationic substitution variants); 3) contains an amino acid substitution at a position normally occupied by cysteine in the BPI sequence (cysteine substitution variants); 4)

contains an amino acid substitution where the substituted amino acid is the amino acid at the corresponding position in the LBP amino acid sequence; and/or 5) contains an insertion or deletion of one or more 5 secondary structure-altering amino acid residues.

Exemplary BPI variants containing a glycosylation site alteration include BPI variants having an amino acid residue other than serine substituted for the serine residue at position 351 of the BPI amino acid sequence.

10 BPI variants of this type are of the formula BPI(S351->X), wherein X is any amino acid other than serine. Preferably, the amino acid substituted at position 351 is alanine. Other BPI variants having a glycosylation site deleted can be generated by, for example, other amino acid substitutions within the glycosylation site.

Additional exemplary BPI variants contain a neutral or anionic amino acid substituted at a cationic residue of the BPI amino acid sequence (cationic 20 substitution variants). For example, one or more of the nonconserved positively-charged residues in BPI (i.e., those residues not found at the corresponding positions in LBP) can be substituted with the corresponding residue or residues in LBP, thus rendering BPI less cationic. 25 Preferably, the cationic substitution variant contains an amino acid substitution in at least one of BPI amino acid residue positions 27, 30, 33, 42, 44, 48, 59, 77, 86, 90, 96, 118, 127, 148, 150, 160, 161, 167, 169, 177, 185, or 198. The cationic substitution variant can contain 30 multiple amino acid substitutions. For example, the cationic substitution variant can contain a neutral or anionic residues at 1) BPI amino acid residue positions 27, 30, 33, 42, 44, 48, and 59; 2) BPI amino acid residue positions 77, 86, 90, 96, 118, and 127; 3) BPI amino acid 35 residue positions 148, 150, 160, 161, 167, 169, 177, 185,

and 198; or 4) BPI amino acid r sidue positions 27, 30, 33, 42, 44, 48, 59, 77, 86, 90, 96, 118, 127, 148, 150, 160, 161, 167, 169, 177, 185, and 198.

Further example BPI variants contain an amino acid substitution at a position normally occupied by cysteine in the BPI sequence (cysteine mutant). The amino acid selected for substitution at this site can be the amino acid in the corresponding position in LBP. For example, a cysteine residue in BPI (which is not conserved in LBP) may be substituted with an alanine residue (the corresponding residue in LBP). Preferably, the amino acid substitution is at a cysteine residue at BPI amino acid residue position 132, 135, or 175. Preferably, alanine or serine is substituted for cysteine. More preferably, alanine is substituted for the cysteine at position 132 of BPI. Cysteine substitution mutants of BPI can prevent aggregation of the resulting RENPs during their production or use.

Another example of a BPI variant includes a BPI

20 variant having an amino acid substitution where the substituted amino acid is the amino acid at the corresponding position in LBP. The amino acid at the corresponding position is determined by aligning the BPI and LBP amino acid sequences so as to maintain the

25 highest level of amino acid sequence identity between the two sequences. For example, an RENP having the formula B_(Q329-S) contains a substitution of the glutamine at BPI residue position 329 with the serine residue at the corresponding LBP residue position 327 (see Figures

30 5A-5F).

Additional exemplary BPI variants contain an insertion or deletion of one or more secondary structure-altering amino acid residues. For example, one or more of the nonconserved proline residues in BPI may

- 26 -

be substituted with the corresponding non-proline r sidue in LBP.

Alternatively, or in addition to the amino acid sequence of BPI and/or a BPI variant, the RENPs can 5 contain an amino acid sequence of LBP, where the LBP amino acid sequence 1) has been altered at a site of glycosylation (e.g., insertion or deletion of a glycosylation site); 2) contains a cationic amino acid substituted at a neutral or anionic amino acid of the LBP 10 amino acid sequence (cationic replacement mutant); 3) contains an amino acid substitution where the substituted amino acid is the amino acid at the corresponding position in the BPI amino acid sequence; and/or 4) contains an insertion or deletion of one or more 15 secondary structure-altering amino acid residues. LBP DNA and amino acid sequence used in the construction of particular RENPs exemplified herein is the amino acid sequence of human LBP in Figs. 5A-B.

Exemplary LBP variants contain a cationic amino 20 acid substituted at a neutral or anionic amino acid of the LBP amino acid sequence (cationic replacement variant). For example, one or more of the nonconserved amino acid residues in LBP (at a position which corresponds to a positively-charged residue in BPI) may 25 be substituted with the corresponding positively-charged residue in BPI, and thus result in an LBP variant having an increased positive charge, thus enhancing binding to the negatively charged phosphate groups in LPS, and/or facilitating interaction with the negatively charged 30 surfaces of Gram-negative bacteria. Positively-charged residues include, by way of example, lysine, arginine, and histidine. Preferably, the substituted cationic amino acid is at least one of LBP amino acid residue positions 77, 86, 96, 118, 126, 147, 148, 158, 159, 161, 35 165, 167, 175, 183, or 196. Cationic replacement

- 27 -

variants can contain multiple amino acid residu substitutions at any combination of the amino acid residues recited above.

Other exemplary LBP variants include an LBP

5 variant having an amino acid substitution where the substituted amino acid is the amino acid at the corresponding position in BPI. For example, L_(A401→D) contains a substitution of the alanine residue of LBP at position 401 with the aspartic acid residue at the 10 corresponding BPI residue position 403.

Further exemplary LBP variants contain an insertion or deletion of one or more one or more secondary structure-altering amino acid residues. For example, one or more of the nonconserved amino acid residues in LBP (at a position which corresponds to a proline in BPI) may be substituted with a proline residue. Preferably, such amino acid alterations alter the secondary structure of the resulting LBP variant so that it is more like the secondary structure of BPI.

Preferably, the RENPs of the invention contain at 20 least one LPS-binding domain of BPI, LBP, a BPI variant, and/or a LBP variant. For example, the LPS-binding domain can be derived from BPI and/or LBP amino acid sequences 17-45, 65-99, and/or 141-167. Preferably, the 25 RENP has an LPS binding affinity that is greater than the LPS binding affinity of LBP, more preferably an LPS binding affinity that is the same or greater than the LPS binding affinity of BPI. Preferably, the RENP has an LPS binding affinity that is about 25-fold to 50-fold, 30 preferably about 50-fold to 100-fold, more preferably about 100-fold to 300-fold greater than the LPS binding affinity of LBP as determined by LPS binding or LPS binding competition assays. The LPS binding affinity of BPI is about 60-fold to 100-fold greater than the LPS 35 binding affinity of LBP.

The RENPs can contain multiple LPS-binding domains derived fr m any of th s LPS-binding proteins. For example, an RENP can be a multivalent chimeric protein (i.e., a fusion protein) composed of an LPS-binding 5 domain of BPI covalently bound (i.e., as in a fusion protein) to an LPS-binding domain of LBP. As used herein, a chimera means a protein comprising all or a portion of a first protein fused to all or a portion of a second protein, which resulting fusion protein may in 10 turn be fused to all or a portion of a third protein. Examples of chimeras include, by way of example, (a) a protein comprising a portion of LBP fused to a portion of BPI, (b) a protein comprising a portion of LBP fused to a portion of BPI which portion of BPI is in turn fused to a 15 portion of an immunoglobulin protein, or (c) a protein comprising a portion of LBP fused to a portion of BPI, which is in turn fused to a portion of LBP. Each protein portion of the chimera may comprise a fragment of the protein, a point mutant of the protein (i.e., a variant), 20 a deletion mutant of the protein, or a point and deletion mutant of the protein.

Examples of BPI fragments which can be incorporated into the RENPs of the invention include the BPI amino acid sequences 1-25, 1-85, 1-137, 1-135, 1-147, 1-159, 88-100, 148-161, 137-199, 44-159, 44-199, 135-199, 100-199, 162-199, 100-147. Examples of LBP fragments which can be incorporated into the RENPs of the invention include LBP amino acid sequences 1-43, 1-87, 26-135, 26-134, 86-99, 101-146, 101-197, 135-197, 137-197, 158-197, 160-197, and/or 147-159. The amino acid sequences of BPI and/or LBP can be comined in any order from N- to C-terminus to provide an RENP having sequences derived from BPI and/or LBP. For example, the RENPs can have the sequences B₁₋₁₃₇L₁₃₇₋₁₉₇, L₁₋₄₃B₄₄₋₁₉₉, B₁₋₁₅₉L₁₅₈₋₁₉₇, B₁₋₃₅L₂₆₋₁₃₅B₁₃₇₋₁₉₉, B₁₋₂₅L₂₆₋₁₃₅B₁₃₇₋₁₉₉, B₁₋₂₅

For example, fusing the N-terminal domain of LBP to the C-terminal domain of BPI results in an RENP which differs from LBP in that the chimera neutralizes endotoxin in whole blood and differs from BPI in that the chimera has a longer circulating half-life in vivo. Such RENPs have significant diagnostic and therapeutic potential. As per the nomenclature described above, RENPs designated BPI-LBP contain all or a part of the N-terminal domain of BPI fused to all or a part of the C-terminal domain of LBP. Likewise, RENPS designated LBP-BPI contain all or a part of the N-terminal domain of LBP fused to all or a part of the C-terminal domain of BPI.

Where the RENP contains amino acid sequences derived from both BPI and LBP, the RENP is preferably 20 composed of a C-terminal fragment of BPI (or a BPI variant) and an N-terminal fragment of LBP (or an LBP variant). Preferably the C-terminal fragment of BPI (or a BPI variant) contains amino acid residues 60-456, 136-456, 199-456, 277-456, 300-456, 200-456, 136-361, 25 136-275, 200-275, or 200-361, more preferably 60-456, more preferably 199-359. The amino acid sequence of BPI from residue 199 to residue 359 contains a region required for neutralizing LPS, i.e., preventing LPS from stimulating an inflammatory response. Preferably, the N-30 -terminal fragment of LBP (or an LBP variant) contains amino acid residues 1-59, 1-134, 1-164, 1-175, 1-274, 1-359, 1-134, or 1-197, more preferably 1-175. addition to the specific amino acid sequences of BPI and LBP recited above, the RENP can also contain amino acid 35 residues derived from the C-terminus of LBP (or an LBP

variant), pr ferably LBP (or LBP variant) amino acid r sidues 360-456 or 274-456.

Polypeptides which bind LPS can be identified using any of several assays well known in the art such as 5 the 1) chromogenic LAL competition assay, 2) binding to LPS immobilized on a surface, and 3) FITC-LPS assay for binding to macrophages. The ability of a polypeptide to neutralize endotoxin can also be determined using methods well known in the art. Endotoxin neutralization assays 10 include assays to examine the ability of a polypeptide to

- 1) prevent LPS-induced TNF release in whole blood,
 - 2) inhibit or prevent TNF production by THP-1 cells,
 - 3) provide protection in a mouse endotoxin challenge assay, and 4) reduce or prevent LPS-induced cytokine
- 15 release and/or mortality in an animal model. Each of these assays are described in detail in the examples section below. The results of the *in vitro* and *in vivo* assays used herein are accepted in the art. The results of these assays are predictive of relevant biological activity *in vivo*, e.g. in humans.

Preferably, the RENPs of the invention have a biological half-life (e.g., serum half-life) which is enhanced relative to the biological half-life of BPI. Preferabl, the half-life of the RENP is enhanced relative to BPI such that the clearance time of the RENP is at least 1.5-fold to 10-fold, preferably about 10-fold to 50-fold, more preferably about 50-fold to 100-fold, even more preferably about 100-fold to 350-fold slower than the clearance rate of BPI. The clearance rate values representing these ranges are from about 8 ml/min to 1.5 ml/min, preferably 1.5 ml/min to 0.26 ml/min, more preferably 0.26 ml/min to 0.13 ml/min, even more preferably about 0.13 ml/min to 0.03 ml/min.

To enhance the RENP half-life, the RENP can be 35 covalently bound to a molecule which enhances the

half-life of the polypeptide. The half-life enhancing molecule can be any of a variety of half-life nhancing molecules. Exemplary half-life enhancing molecules include immunoglobulin fragments, a half-life determining 5 portion of LBP, a half-life determining portion of an LBP variant, or polyethylene glycol (PEG), preferably a half-life determining portion of LBP or an LBP variant. Preferably, where the half-life enhancing molecule is a portion of LBP or an LBP variant, the half-life enhancing 10 molecule is derived from the N-terminus of the LBP or LBP variant amino acid sequence, more preferably from amino acid residues 1-59, 1-134, 1-274, 1-359, 1-134, 1-164, 1-175, or 1-197, most preferably 1-164 or 1-175. Methods of attachment of PEG moieties to a protein (i.e., 15 PEGylation) are well known in the art and are exemplified in U.S. patent nos. 4,179,337; 5,166,322; 5,206,344; and PCT application serial no. PCT/US94/11624, published April 28, 1995.

As used herein, an RENP-Ig chimeric protein is an 20 RENP which (i) contains a portion of BPI or LBP (at least 10 amino acid residues in length of (a) BPI, or (b) BPI variant, or (c) LBP, and/or (d) LBP variant) fused at the C-terminus to the N-terminus the Fc portion of an immunoglobulin molecule, and (ii) is capable of (a) 25 binding to LPS, (b) competing with BPI or LBP for binding to LPS, and (c) inhibiting the production of $TNF\alpha$ by human monocytes. For example, the portion of the immunoglobulin molecule is derived from an IgG molecule, specifically from an IgG₁ heavy chain Fc domain. RENP-Ig 30 chimera is a fusion protein composed predominantly of sequences derived from BPI, variant BPI, LBP and/or variant LBP. The term "LBP-BPI-IgG chimera" indicates that the RENP-Ig chimera contains amino acid sequences derived from both BPI (or a BPI variant) and LBP (or an 35 LBP variant).

.- 32 -

Identification of a half-life enhancing polypeptide sequenc (.g., a polypeptide derived from an immunoglobulin, LBP, or LBP variant) can be accomplished using methods well known in the art. For example, the 5 test polypeptide with and without the half-life enhancing molecule bound to it are injected into an animal model to determine the effects of the putative half-life enhancing molecule. If the half-life of the polypeptide with the molecule is enhanced relative to the half-life of the 10 polypeptide without the molecule, then the molecule is a half-life enhancing molecule suitable for use in the RENPs of the invention. For example, a putative half-life enhancing amino acid sequence is incorporated into a fusion protein with BPI. Both native BPI and the 15 BPI fusion protein are injected into mice. If the BPI fusion protein has a half-life significantly greater than the half-life of native BPI, then the amino acid sequence in the BPI fusion has half-life enhancing characteristics, and thus can be incorporated into the 20 RENPs of the invention.

<u>Vectors</u> and constructs

Any nucleic acid vector can be used to express DNA encoding an RENP of the invention. The vectors containing the DNA sequence (or the corresponding RNA sequence) which may be used in accordance with the invention may be any prokaryotic or eukaryotic expression vector containing the DNA (e.g., cDNA) or the RNA sequence of interest. A variety of suitable vectors are publicly available and well known in the art. For example, a plasmid can be cleaved to provide linear DNA having ligatable termini. These termini are bound to exogenous DNA having complementary, like ligatable termini to provide a biologically functional recombinant

- 33 -

DNA molecule having an intact replicon and a desired phenotypic property.

A variety of techniques are available for DNA recombination in which adjoining ends of separate DNA 5 fragments are tailored to facilitate ligation. vector is constructed using known techniques to obtain a transformed cell capable of expression of the RENP. transformed cell is obtained by contacting a target cell with a RNA- or DNA-containing formulation permitting 10 transfer and uptake of the RNA or DNA into the target cell. Such formulations include, for example, plasmids, viruses, liposomal formulations, or plasmids complexed with polycationic substances such as poly-L-lysine or DEAC-dextran, and targeting ligands. Transformed cells 15 containing a construct encoding an RENP of the invention are also known in the art as "host vector systems". Vectors for use in the construction of constructs. encoding the RENPs of the invention, as well as methods for molecular cloning, nucleic acid manipulation, and 20 transformation of both eukaryotic and prokaryotic host cells are well known in the art (see, for example, Sambrook et al. Molecular Cloning: A Laboratory Manual, 2nd Ed., 1989, Cold Spring Harbor Laboratory Press, Cold Spring Harbor, NY; hereby incorporated by reference with 25 respect to bacterial and eukaryotic vectors, and methods and compositions for molecular cloning, nucleic acid manipulation, and transformation techniques).

The constructs of the invention may include promoter sequences to enhance expression of the 30 RENP-encoding DNA, as well as other sequences (e.g., enhancers) which facilitate or enhance DNA expression. In addition, the RENP-encoding constructs can contain other components such as a marker (e.g., an antibiotic resistance gene (such as an ampicillin resistance gene) or β-galactosidase) to aid in selection of cells

containing and/or xpressing the construct, an origin of replication for stable r plication of the construct in a bacterial cell (preferably, a high copy number origin of replication), a nuclear localization signal, or other elements which facilitate production of the DNA construct, the protein encoded thereby, or both.

In general, the RENPs of the invention are constructed from a DNA sequence encoding BPI, a BPI variant, LBP, an LBP variant, as well as various

10 half-life enhancing molecules known in the art such as immunoglobulin fragments. Both BPI and LBP have been cloned and their DNA and amino acid sequences determined (Figures 3A-3B and 4A-4B, respectively). The DNA and amino acid sequences of numerous immunoglobulins are

15 known in the art. For example, the DNA sequence of IgG, IgG_{2a}, and IgG₄ are suitable for use to enhance the half-life of the RENPs of the invention.

Expression of recombinant endotoxin-neutralizing polypeptides

DNA or RNA sequences in a host cell are known in the art (see, for example, Sambrook et al., supra; hereby incorporated by reference with respect to methods and compositions for eukaryotic and prokaryotic expression of a DNA encoding an RENP). Where the transformed cell is a prokaryotic host cell, the preferred host is Escherichia coli. Where the transformed cell is a eukaryotic host cell, preferably the host cell is a mammalian cell or a yeast cell. Preferably, the mammalian host cell is a 30 Chinese Hamster Ovary (CHO) cell. Preferably a strain of Pichia pastoris.

For prokaryotic expression, the construct should contain at a minimum a bacterial origin of replication

- 35 -

and a bacterial promoter op rably linked to the RENP-encoding DNA. For eukaryotic expression, the construct should contain at a minimum a eukaryotic promoter operably linked to a DNA of interest, which is 5 in turn operably linked to a polyadenylation sequence. The polyadenylation signal sequence may be selected from any of a variety of polyadenylation signal sequences known in the art. Preferably, the polyadenylation signal sequence is the SV40 early polyadenylation signal 10 sequence. The eukaryotic construct may also include one or more introns, which can increase levels of expression of the DNA of interest, particularly where the DNA of interest is a cDNA (e.g., contains no introns of the naturally-occurring sequence). Any of a variety of 15 introns known in the art may be used. Preferably, the intron is the human β -globin intron and inserted in the construct at a position 5' to the DNA of interest.

Purification of RENPs

Purification of the RENPs of the invention can be
20 performed according to any of a variety of protein
purification techniques known in the art including gel
electrophoresis, immunoprecipitation, ion exchange
chromatography, affinity chromatography, or combinations
thereof (see, for example, <u>Guide to Protein Purification</u>,
25 Deutscher, ed., Academic Press, Inc., San Diego, CA,
1990). Preferably, purification of RENPs is accomplished
by a combination of column chromatographic techniques.
For example, RENPs can be purified using a four-step
purification procedure using 1) a cation exchange column
30 (e.g., CM Sepharose), 2) an anion exchange column (e.g.,
Fast Q Sepharose), 3) a second cation exchange column
(e.g., CM Sepharose), and 4) a gel filtration sizing
column (e.g., Sepharose CL6B).

- 36 -

Pharmaceutical compositions

Th RENPs f the inv ntion can be formulated as an active ingredient in a pharmaceutical composition. general, the pharmaceutical composition contains a 5 therapeutically effective amount of an RENP and a pharmaceutically acceptable carrier. The pharmaceutical composition can contain one or more RENPs. The amount of RENP which constitutes a therapeutically effective amount will vary according to the time of administration (e.g., 10 therapeutic or prophylactic administration), the disease or condition to be treated, the route of administration, and various patient-dependent factors such as age, weight, gender, and severity of disease. Specific therapeutically effective amounts appropriate for 15 administration are readily determined by one of ordinary skill in the art (see, for example, Remington's Pharmaceutical Sciences, 18th ed., Gennaro, ed., Mack Publishing Company, Easton, PA, 1990).

Pharmaceutically acceptable carriers suitable for use in the RENP-containing pharmaceutical compositions of the invention are well known to those skilled in the art. Selection of the pharmaceutically acceptable carrier will depend upon a variety of factors including the RENP to be administered, the route of administration, and the condition to be treated.

Pharmaceutically acceptable carriers suitable for use with the RENPs of the invention include, but are not limited to, 0.01-0.1 M and preferably 0.05 M succinate buffer or 0.8% saline. Additionally, such

30 pharmaceutically acceptable carriers may be aqueous or non-aqueous solutions, suspensions, and emulsions.

Further, pharmaceutically acceptable carriers may include detergents, phospholipids, fatty acids, or other lipid carriers. Examples of non-aqueous solvents are propylene glycol, p lyethyl ne glycol, vegetable oils such as olive

- 37 -

oil, and injectable organic sters such as ethyl oleate.
Aqueous carriers include water, alcoholic/aqueous
solutions, emulsions or suspensions, including saline and
buffered media. Parenteral vehicles include sodium
5 chloride solution, Ringer's dextrose, dextrose and sodium
chloride, lactated Ringer's or fixed oils.

Pharmaceutically acceptable carriers for use with the RENPs of the invention include lipid carriers. A lipid carrier is any lipid-soluble substance which 10 inhibits protein precipitation and in which the proteins of the subject invention are soluble. Lipid carriers can be in the form of sterile solutions or gels, or can be detergents or detergent-containing biological surfactants. Examples of nonionic detergents include 15 polysorbate 80 (also known as TWEEN 80 or polyoxyethylenesorbitan monooleate). Examples of ionic detergents include, but are not limited to, alykltrimethylammonium bromide. Exemplary lipid carriers and methods for solubilizing BPI, and thus which can be 20 used in pharmaceutical compositions containing an RENP of the invention, are described in USPN 5,234,912, incorporated herein by reference.

Where the pharmaceutically acceptable carrier is a lipid carrier, the lipid carrier may be a liposome. A liposome is any phospholipid membrane-bound vesicle capable of containing a desired substance, such as BPI or BPI variant, in its hydrophilic interior. Intravenous vehicles include fluid and nutrient replenishers, electrolyte replenishers such as those based on Ringer's dextrose, and the like. Preservatives, other pharmaceutically active compounds, and other additives may also be present, such as, for example, antimicrobials, antioxidants, chelating agents, inert gases and the like.

Disease conditions amenable to treatment with RENPs

Various diseas c nditions are amenable to treatment using the recombinant LPS-neutralizing proteins of the invention. In general, any condition of a mammalian subject (e.g., human, canine, feline, or bovine, preferably a human) which is associated with a toxic effect of endotoxin can be treated by administration of the RENPs of the invention. Endotoxin-related disorders amenable to treatment

- include, but are not limited to, endotoxin-related shock, endotoxin-related disseminated intravascular coagulation, endotoxin-related anemia, endotoxin-related thrombocytopenia, endotoxin-related adult respiratory distress syndrome, endotoxin-related renal failure,
- endotoxin-related liver disease or hepatitis, systemic immune response syndrome (SIRS) resulting from Gram-negative infection, Gram-negative neonatal sepsis, Gram-negative meningitis, Gram-negative pneumonia, neutropenia and/or leucopenia resulting from
- 20 Gram-negative infection, hemodynamic shock and endotoxin-related pyresis. Endotoxin-related pyresis is associated with certain medical procedures, such as, for example, trans-urethral resection of the prostate, and gingival surgery. The presence of endotoxin may result
- 25 from infection at any site with a Gram-negative organism, or conditions which may cause ischemia of the gastrointestinal tract, such as hemorrhage, or surgical procedures requiring extracorporal circulation. The important role of endotoxin in hemorrhage (with
- 30 endogenous LPS translocation from the gut), trauma, and sepsis is well known. One skilled in the art can recognize additional conditions which can be treated using the therapy of the invention.

The recombinant, endotoxin-neutralizing proteins
35 of the invention can also be administered to a patient

prophylactically, e.g. to a patient at risk of an endotoxin-related disorder. For example, the RENPs can be administered to a patient who has a Gram-negative infection and is at risk of bacteremia, but who has not yet exhibited symptoms associated with the toxic effects of endotoxin. The RENPs can also be administered prior to surgery where the risk of introduction of endotoxin into the patient is substantial. One of ordinary skill in the art can readily recognize other instances in which prophylactic administration of a RENP is appropriate. The conditions which identify an individual as being at risk of an endotoxin-related disorder are well known in the art.

Administration of RENPs

15 The recombinant, LPS-binding protein of the invention may be administered using various methods well known in the art. U.S. Pat. Nos. 5,171,739; 5,308,834; and 5,334,584; each incorporated herein by reference, describe methods and compositions for administration of 20 BPI, and thus can provide additional guidance for administration of the RENPs of the invention. For example, the recombinant, LPS-binding protein can be administered by injection or inhalation. Administration by injection can be an intravenous, intramuscular, or 25 subcutaneous route, or by direct injection directly into a site of infection (e.g., tissue or body cavity). Preferably, injection is intravenous. Administration by inhalation is accomplished by delivery of the RENP to the lungs via an aerosol delivery system or via direct 30 instillation. The aerosol may be nebulized. Various devices and methods for aerosol drug delivery are well known in the art. Methods for determining the appropriate route of administration and dosage are generally determined on a case-by-case basis by the

attending physician. Such determinations ar routine to one f ordinary skill in the art (see, for xample, Remington's Pharmaceutical Sciences, 18th ed., Gennaro, ed., Mack Publishing Company, Easton, PA, 1990).

5 Therapeutically effective amounts of an RENP can be determined according to methods well known to those skilled in the art. Specific dosages will vary according to a variety of factors, including the time of administration (e.g., therapeutic or prophylactic 10 administration), the disease or condition to be treated, the route of administration, the RENP to be administered, and various patient-dependent factors such as age, weight, gender, and severity of disease. The specific dosage appropriate for administration is readily 15 determined by one of ordinary skill in the art according to the factors discussed above (see, for example, Remington's Pharmaceutical Sciences, 18th ed., Gennaro, ed., Mack Publishing Company, Easton, PA, 1990). addition, the estimates for appropriate dosages in humans 20 may be extrapolated from determinations of the in vitro LPS binding affinity of the RENP used, the amount of the RENP effective to inhibit cytokine production by mononuclear cells in vitro, the amount of RENP effective to provide protection to LPS challenge, and/or various 25 other in vitro and in vivo assays indicative of the biological activity of the RENP.

In general, the amount of RENP administered is an amount effective to bind LPS and thereby inhibit the undesirable biological activities associated with LPS including monocyte and neutrophil activation, TNF production, cytokine production, and other biological phenomena triggered by LPS in endotoxin-related disorders. Preferably, the amount of RENP administered is an amount effective to bind LBP and inhibit LPS-mediat d stimulation of neutrophils and mononuclear cells.

- 41 -

In therapeutic administration of the RENPs of the inventi n, an eff ctiv amount of an RENP is an amount effective to bind to LPS and thereby inhibit LPS-mediated stimulation of neutrophils and mononuclear cells in a 5 subject having an endotoxin-related disorder. As used herein, "inhibit" means to inhibit at a level which is statistically significant and dose dependent. "statistically significant" and "dose dependent" are well known to those skilled in the art. In general, an 10 effective amount of an RENP in a pharmaceutical composition for treatment of a patient having an endotoxin-related disorder is an amount sufficient to deliver to the subject a recombinant protein of the subject invention at a concentration of between about 15 0.1 mg/kg of body weight and about 100 mg/kg of body weight, preferably between about 1 mg/kg of body weight and about 10 mg/kg of body weight. Preferably, the RENP(s) is administered by injection, infusion, or as an injected bolus so as to maintain a circulating RENP 20 concentration of about 1-10 μ g/ml. The preferred circulating RENP concentration can vary according to a variety of factors, including the LPS binding affinity of the specific RENP(s) administered.

As used herein, a prophylactically effective

25 amount of an RENP in a pharmaceutical composition for the
prevention of an endotoxin-related disorder is an amount
effective to bind LPS and prevent LPS-mediated biological
activity, e.g., LPS-mediated stimulation of monocytes and
neutrophils. In general, a prophylactically effective

30 amount of an RENP is an amount of a composition effective
to deliver between about 0.1 mg/kg of body weight and
about 100 mg/kg of body weight, preferably between about
1 mg/kg of body weight and about 10 mg/kg of body weight,
to the patient at risk of an endotoxin-related disorder.

- 42 -

The invention also provides an article of manufacture comprising packaging material and a pharmaceutical composition contained within the packaging material. The packaging material includes a label which indicates that the pharmaceutical composition can be used for treating a subject suffering from an endotoxin-related disorder and/or for preventing an endotoxin-related disorder (e.g., inflammation) in a subject. The pharmaceutical composition contains a therapeutically effective and/or prophylactically effective amount of an RENP and a pharmaceutically acceptable carrier.

Assessment of therapy

The efficacy of the therapeutic or prophylactic 15 use of the RENPs of the invention can be determined by monitoring patient symptoms associated with an endotoxin-related disorder. Such symptoms, and methods for monitoring, are well known in the art. For example, where the RENP is used in the treatment of a patient 20 having an endotoxin-related disorder, the effectiveness of the RENP therapy can be assessed by monitoring fever, blood pressure, cytokine levels, and/or LPS levels in the patient's blood stream. The presence of LPS in the blood stream can be assayed as described above. Where the 25 patient is not responding, it may be desirable to increase the dosage of the RENP pharmaceutical composition or, where the patient is not responding favorably, discontinue the RENP regimen.

Detectably-labeled RENPs

Various detectable labels, as well as methods of attachment of such labels to a protein, are well known in the art. Detectable labels can be any molecule recognized in the art as a means for identifying and/or

detecting a protein to which the detectable label is bound. Exemplary "detectable labels" include, but ar not limited to radionucleotides, fluorescent moieties, biotin, and antigenic molecules (e.g., a polypeptide 5 which can be specifically bound by an anti-polypeptide antibody). Thus, detectable labels include a portion of a chimeric protein (e.g., a fusion protein or genetically engineered protein) where a portion of the chimeric protein can be detected by, for example, binding of a 10 detectably labeled antibody or other detectably labeled molecule which specifically binds the chimeric protein portion. For example, where the RENP contains a portion of the amino acid sequence of BPI, and an antibody which specifically binds that amino acid sequence of BPI in the 15 context of the RENP is available, the BPI amino acid sequence is the detectable label.

Methods for attaching (e.g., covalently binding) a detectable label to a protein are well known in the art. For example, methods for preparation of 125I-labeled 20 proteins, biotin-labeled proteins, and FITC-labeled proteins are well known. Methods for detectably labeling antibodies are also well known in the art. Methods for the production of antibodies for use in the subject invention (e.g., anti-BPI, anti-LBP, anti-BPI variant, anti-LBP variant, and anti-immunoglobulin fragment antibodies) are well known in the art (see, for example, Antibodies: A Laboratory Manual, Harlow and Lane, eds., Cold Spring Harbor Laboratory, Cold Spring Harbor, NY, 1988).

30 Detection of LPS in vitro

The detectably labeled RENPs of the invention can be used in various methods for the detection of LPS either in vitro or in vivo. Samples for which in vitro LPS detection is desirable include samples from a patient

suspected of having a Gram-negative infection, and samples from a pr duct for use in a medical application (e.g., a recombinant protein solution where the protein was expressed in E. coli). Patient samples include samples of any body fluid, preferably blood or urine. Samples may be pre-treated prior to testing by, for example, concentrating the sample, or centrifugation to remove cells and cellular debris.

In general, in vitro detection of LPS in a sample suspected of containing LPS (test sample) is performed by contacting the test sample with an RENP of the invention for a time sufficient for the formation of RENP-LPS complexes, and the RENP-LPS complexes detected. The RENP-LPS complexes can be detected by virtue of a detectable label attached to the RENP, or by the binding of an anti-LPS antibody. Binding of the anti-LPS antibody can subsequently be detected by virtue of a detectable label bound to the antibody, or by the binding of a detectably labeled anti-anti-LPS antibody to the RENP-LPS-antibody complex.

The in vitro assay can be performed in solution by mixing the sample with a solution containing RENP, separation of RENP-LPS complexes (e.g., by immunoprecipitation), and detection of the RENP-LPS

25 complexes formed, e.g., by virtue of a detectable label bound to the RENP. Alternatively, the in vitro assay is performed with RENP bound to a support, e.g., a polymeric substrate such as a microtiter well or a latex bead.

Methods for binding proteins to a support are well known in the art. For example, an anti-RENP antibody can be bound to the support and the RENP subsequently bound to the support via binding to the anti-RENP antibody. After binding of the RENP to the support, the sample is then contacted with the support-bound RENP and any LPS in the sample allowed to bind to the RENP. Unbound material is

- 45 -

then washed away, and the RENP-LPS complexes detected by the binding f detectably label d RENP or d tectably labeled anti-LPS antibody.

The in vitro assay can also be performed as a

5 competition binding assay. For example, a sample
suspected of containing LPS (test sample) and a known
amount of detectably labeled RENP are incubated together
with a support having LPS bound to its surface. The test
sample and the RENP may be preincubated prior to contact

10 with the support-bound RENP. The level of detectably
labeled RENP bound to the support in the test sample is
compared to the level of detectably labeled RENP bound to
the support in a negative control sample (detectably
labeled RENP alone). A level of binding of detectably

15 labeled RENP in the test sample which is lower than
binding of detectably labeled RENP in the negative
control sample is indicative of the presence of LPS in
the sample.

In an alternative embodiment, the competition

20 binding assay is performed with support-bound RENP. In
this latter assay, detectably labeled LPS (e.g.,
radiolabeled LPS) is mixed with the test sample suspected
of containing LPS, and the samples contacted with the
support-bound RENP, and the amount of detectably labeled

25 LPS bound to the support bound RENP detected. A level of
detectably labeled LPS bound to the support in the test
sample which is significantly lower than the amount of
detectably labeled LPS in the negative control sample
(radiolabeled LPS alone) is indicative of the presence of
30 LPS in the test sample.

As is apparent from the description above, the in vitro LPS assays of the invention can be performed both qualitatively and quantitatively. For example, quantitative assays can be performed by comparing the results obtained with the test sample to results obtained

with parallel samples containing known amounts of LPS.

Quantitative in vitro assays ar indicative of, f r
example, the severity of Gram-negative infection in a
patient sample from whom the sample was obtained, or a

5 degree of contamination where the test sample is a fluid
for administration to a patient (e.g., where the assay is
performed as a step in quality control). One of ordinary
skill in the art will appreciate upon reading the
above-described in vitro assays that numerous variations
10 of these assays can be performed without departing from
the spirit or the scope of the invention.

Detection of LPS in vivo

Detectably labeled RENPs of the invention, preferably RENPs having an increased LPS binding affinity relative to LBP, can be used as a diagnostic to identify a site of Gram-negative bacterial infection in a patient. For example, a detectably labeled RENP is administered to a patient suspected of having a Gram-negative infection. Preferably, the detectable label is a radionucleotide such as ¹²³I, ¹²⁴I, ¹²⁵I, ¹²⁹I, ¹³¹I, or other beta-emitting radionuclide which can be readily detected with either a hand-held gamma radiation detection device or by nuclear medicine scan. Alternatively, the detectable label is a fluorescent molecule or other visually detectable label which can be visualized during, for example, endoscopy. Detection can be facilitated by increasing the ratio of detectable label to RENP.

The detectably labeled RENP is administered to the patient in an amount sufficient for binding of the RENP 30 to the suspected infection site and detection of the detectable label. The detectably labeled RENP can be administered by injection, preferably by either intravenous injection or by direct injection into the body cavity or tissue suspected of containing the

- 47 -

infection site. In general, the amount of det ctably labeled RENP administered will vary with according to numerous variables including the RENP and detectable label used, the location of the suspected site of infection, the route of administration, and various patient factors including size, weight, age, and suspected severity of the disease.

is allowed to circulate to reach the site of infection
and/or incubate over the suspected site of infection.
Bound detectably labeled RENP is detected using methods appropriate for the label used. For example, where the detectable label is a radionucleotide, bound RENP is detected using a radiation detecting device. Using this method, the site and the extent of a Gram-negative infection can be determined. Where desirable, the detectably labeled RENPs can be used to label a site or sites of infection which can then be imaged using any of a variety of imaging techniques known in the art (e.g., 20 X-ray, CAT scan, MRI, or PET scan).

LPS decontamination using RENPs

The RENPs of the invention can also be used in the decontamination of a product prior to its medical application. For example, where a recombinant protein 25 has been produced by expression in E. coli, a solution containing the recombinant protein can be applied to a support having bound RENP (e.g., an affinity column). LPS in the solution binds to the RENP bound to the support, and the LPS-free solution is collected. If necessary, the decontamination step can be repeated multiple times until an acceptably low amount of LPS (e.g. 0 to 0.001 ng/ml is detected in the solution. Such decontamination procedures using the RENPS of the

- 48 -

invention can be used as a final step in quality control of, f r example, recombinantly produced pharmaceuticals.

EXAMPLES

The following examples are put forth so as to

5 provide those of ordinary skill in the art with a
complete disclosure and description of how to carry out
the invention and is not intended to limit the scope of
what the inventors regard as their invention. Efforts
have been made to ensure accuracy with respect to numbers

10 used (e.g., amounts, temperatures, etc.), but some
experimental error and deviation should be accounted for.
Unless indicated otherwise, parts are parts by weight,
molecular weight is weight average molecular weight,
temperature is in degrees Centigrade, and pressure is at

15 or near atmospheric.

Example 1: Construction of RENPs

Specific examples of RENPs are described in Table 3, and are additionally designated by a construct name (e.g., NCY103) or lot number of the protein stock.

Table 3

		xamples of REN	<u>Ps</u>
	SEQUENCE	CONSTRUCT NAME OR LOT #	DESCRIPTION
5	BPI	NCY101	Native sequence
	L ₁₋₁₉₇ (148->V) ^B 200-456(N208-> D)	NCY103	LBP-BPI chimera
	B ₁₋₁₉₉ L ₂₀₀₋₄₅₆	NCY104	BPI-LBP chimera
	B(S351->A)	NCY105	Glycosylation site deleted
10	B(DS200->DP)	NCY106	Formic acid cleavage site inserted
	L ₁₋₁₉₉ B ₂₀₀₋₄₅₆ (S351->A)	NCY107	LBP-BPI chimera with glycosylation site deleted
	B ₁₋₁₉₉	NCY108	N-terminal domain of BPI
	B ₍₁₋₁₉₀₎	Lot #159699	N-terminal BPI fragment
	B ₍₁₋₂₃₆₎	Lot #159695	N-terminal BPI fragment
15	B ₍₁₋₂₁₂₎	Lot #159693	N-terminal BPI fragment
	B ₁₋₁₉₉ Fc	NCY110	N-terminal BPI-IgG chimera
	B ₂₀₀₋₄₅₆	NCY112	C-terminal fragment of BPI
	L ₁₋₅₉ B ₆₀₋₄₅₆	NCY114	LBP-BPI chimera
	L ₁₋₁₃₄ B ₁₃₅₋₄₅₆	NCY115	LBP-BPI chimera
20	L ₁₋₂₇₅ B ₂₇₈₋₄₅₆	NCY116	LBP-BPI chimera
	L ₁₋₃₅₉ B ₃₆₀₋₄₅₆	NCY117	LBP-BPI chimera
	L ₍₁₋₁₆₄₎ B ₍₂₀₀₋₄₅₆₎	Lot #164325	LBP-BPI chimera
	L ₍₁₋₁₇₅₎ B ₍₂₀₀₋₄₅₆₎	Lot #164326	LBP-BPI chimera
	L ₁₋₁₉₇ B ₂₀₀₋₄₅₈	NCY118	LBP-BPI chimera
25	B _(P61->C)	NCY119	Cysteine insertion
	B(C132->A)	NCY120	Cysteine substitution

	B _(C132->S)	NCY121	Cysteine substitution
	B(C1\$5->S)	NCY122	Cysteine substitution
	B _(C176->S)	NCY123	Cysteine substitution
	T	<u>table 3</u> (continued)	
5	SEQUENCE	CONSTRUCT NAME OR LOT #	DESCRIPTION
	B(C132->A)(C135->S)(C175->S)	NCY124	Multiple cysteine substitution
	B _(1-132->A) (C135->S)(C175->S)	NCY125	Multiple cysteine substitution
	L ₍₁₋₁₃₄₎ B ₍₁₃₆₋₃₆₁₎ L ₍₃₆₀₋₄₅₆₎	NCY133	LBP-BPI chimera
	L ₍₁₋₁₃₄₎ B ₍₁₃₆₋₂₇₅₎ L ₍₂₇₄₋₄₅₆₎	NCY134	LBP-BPI chimera
10	L ₍₁₋₁₉₈₎ B ₍₂₀₂₋₂₇₅₎ L ₍₂₇₄₋₄₅₆₎	NCY135	LBP-BPI chimera
	L ₍₁₋₁₉₈₎ B ₍₂₀₂₋₃₆₁₎ L ₍₃₆₀₋₄₅₆₎	NCY136	LBP-BPI chimera
	B ₍₁₋₄₁₎ L ₍₄₂₋₁₉₉₎ B ₍₂₀₀₋₄₅₆₎	Lot #162303	BPI-LBP-BPI chimera
•	B ₍₁₋₁₉₀₎ (C173→A)	Lot #162305	N-terminal BPI fragment with cationic substitution
15	B _{(K27} ->S)(K30->L)(K33->T) (K42->R)(K44->P)(K48->R)(R59- >H) (B _{CAT7})	NCY137	Cationic Substit. (7)
20	B _(K77->S) (K86->R)(K90->R) (R96->S)(K118->L)(K127->R) (B _{CA76})	NCY138	Cationic Substit. (6)
	B(K148->G)(K150->D)(K160->N) (K161->Q)(R167->Q)(K169->V) (K177->M)(K185->D)(K198->E) (B _{CAT9})	NCY139	Cationic Substit. (9)
30	$\begin{array}{l} \mathbf{B}_{(K77->S)(K86->R)(K90>R)} \\ (K96->S)(K118->L)(K127->R)(K148->G) \\ (K160->D)(K160->N)(K161->Q)(R167->Q)(K169->V)(K177->M)(K185->D)(K198->E) \\ (\mathbf{B}_{CAT15}) \end{array}$	NCY140	Cationic Substit. (15)
	L(S77->K)(R86->K)(R90- >K)(S96->K) (L118->K)(R126->K) (L _{TAT6})	NCY141	Cationic Repl. (6)

	L(G147->K)(D148->K)(N158->K) (Q159->K)(Q165->R)(V167->K)(M 175->K)(D185->K)(E196->K) (L _{CAT9})	NCY142	Cationic Repl.(9)
5 10	L(S77->K)(R86->K)(R90- K)(S96->K) (L118->K)(R126->K)(G147->K)(D 148->K)(N158->K)(Q159->K)(Q16 5->R)(V167->K)(M175->K)(D185- >K)(E196->K) (L	NCY143	Cationic Repl. (15)
į	L ₍₁₋₁₉₈₎ B ₍₂₀₁₋₄₅₆₎ Pc	NCY144	LBP-BPI-IgG chimera

		Table 3 (contin	nued)
	LBP	NCY102	native sequence
	L ₁₋₁₉₉	NCY109	N-terminal LBP fragment
	L ₁₋₁₉₉ Fc	NCY111	LBP-1g chimera
5	L ₂₀₀₋₄₅₈	NCY113	C-terminal LBP fragment
	L(A132-C)	NCY126	Cysteine insertion
10	L(C81→F)	NCY127	Cysteine substitution
	L(C81S)	NCY128	Cysteine substitution
	L _(C135→S)	NCY129	Cysteine substitution
	L _(175→S)	NCY130	Cysteine substitution
	^L (C81→F)(C135→S)(C175→S)	NCY131	Multiple cysteine substitution
	L(C81→S)(C135→S)(C175→S)	ису132	Multiple cysteine substitution

The proteins encoded by the LBP and L₁₋₃₅₉B₃₆₀₋₄₅₆ constructs facilitated the LPS-mediated cellular
15 response, indicating that LBP amino acid residues 275-359 are required for this LBP activity.

The cDNA sequences of BPI and LBP are shown in Figures 3A-3D and 4A-C, respectively, with each nucleotide designated numerically. DNA encoding the 20 RENPs can be prepared using a variety of techniques well known in the art, including protein fusion techniques, site-directed mutagenesis, and PCR (see, for example, Sambrook et al., supra; Zoller, M.J., et al., Methods Enzymol. 154:329 (1987)). For example, in the 25 construction of the RENP L₁₋₁₉₇B₂₀₀₋₄₅₆, the sequence "ATAGAT₇₂₃" and "ATTGAC₇₀₀" was chosen as a convenient site to insert a ClaI restriction site (ATCGAT) by which to recombine portions of both BPI (former) and LBP (latter). Oligonucleotide primers were designed which overlap this region but contain the ClaI sequence, and

- 53 -

w re synthesized n an ABI 380B synthesizer (Appli d Biosystems Inc., F ster City, CA). Additional primers were designed to bind to the 5' and 3'- ends of both molecules, which primers contained NheI (5') and XhoI (3') restriction sites for insertion into the vector. These primers were used to amplify portions of the cDNA molecules encoding amino acid residues 1-199 (A) and 200-456 (B) of LBP and BPI by cyclic DNA amplification. The resulting DNA fragments were digested with the appropriate restriction enzymes and then purified by gel electrophoresis.

Example 2: Mammalian Expression

In order to produce BPI, LBP, or RENPs of the invention in mammalian cells, the cDNA sequences were 15 inserted into a suitable plasmid vector. A suitable vector for such an application is pSE, which contains the origin of replication and early and late promoters of SV40, followed by multiple insert cloning sites, followed by the termination sequences from the hepatitis B surface 20 antigen gene. An origin of bacterial DNA replication, and the genes encoding ampicillin resistance and dihydrofolate reductase were also included in the plasmid for production of large amounts of DNA using bacterial host cells. Similar vectors have been used to express 25 other foreign genes (Simonsen et al., Biologicals 22:85 (1994). Another suitable vector, particularly for rapidly obtaining small quantities of RENPs was pCIP4 (Invitrogen Corp., San Diego, California). pCEP4 contains a CMV promoter, followed by multiple insert 30 cloning sites, followed by SV40 termination sequences. Also contained within the plasmid are an origin of bacterial DNA replication, and the genes encoding resistance to ampicillin and hygromycin B. With pCEP4 and pSE, the same insert cloning sites as pSE for easy

insert shuttling between the vectors were used. Once introduced into mammalian cell hosts, this specialized plasmid replicates as an episome, allowing semistable amplification of introduced DNA sequences. The high gene copy number is maintained through the selective pressure of culture in the presence of hygromycin B.

A second expression system (EBV/293) was used to rapidly obtain small quantities of recombinant proteins of the subject invention when useful. This system was constructed to use the same insert cloning sites as pSE for easy insert shuttling, but utilized the Epstein-Barr virus promoter (EBV) to drive heterologous expression (pCEP4). Once introduced into mammalian cell hosts, this specialized plasmid replicates as an episome, allowing semistable amplification of introduced DNA sequences. The high gene copy number is maintained through the selective pressure of culture in the presence of hygromycin plus G418. Similar expression systems are commercially available (e.g., Invitrogen, Inc., San Diego, CA).

Vector DNA was prepared for acceptance of BPI cDNA by digestion with Nhe I and Xho I, and was subsequently dephosphorylated by treatment with alkaline phosphatase. The prepared fragments encoding BPI, LBP, or an RENP were ligated into pSE or pCEP4, and the resulting recombinant colonies were screened by agarose gel electrophoresis. Subsequently, the DNA sequences were confirmed by standard enzymatic sequencing methods (e.g., Sanger, 1974).

gradients with Plasmid or Midi Kits (Qiagen, Chatsworth, California) was used to transform Chinese hamster ovary strain DUXB11 (pSE) and 293-EBNA cells (Invitrogen Corp., San Diego, California) (pCEP4). Transfection was performed using lipofectin (Bethesda, Research Labs,

Gaithersberg, MD) by standard methods. The resulting transformed cells were select d in GHT minus medium (DUKXB11s) or in REM and 10% calf serum (293s). For the DUKXB11s, clones were selected and were passed through sequential rounds of culture in increasing concentrations of methotrexate in order to amplify the DHFR gene and associated heterologous genes. Supernatants from transfected cells, either mixed populations or clones derived from the mixed population, were assayed for RENPs by ELISA using antibodies specific for BPI, LBP, or immunoglobulin as appropriate.

Example 3: Yeast Expression

BPI and L₁₋₁₉₇B₂₀₀₋₄₅₆ were successfully expressed in the methylotrophic yeast *Pichia pastoris*. Pichia was chosen as a suitable expression system for BPI and RENPs due to its lack of LPS (endotoxin to which BPI and RENPs bind) and its ability to produce high levels of mammalian proteins.

Pichia pastoris strain GS115 (Invitrogen, San 20 Diego, California) was transformed with plasmids encoding BPI and $L_{1-197}B_{200-456}$, and transformed colonies were selected according to the procedures outlined by Invitrogen (A Manual of Methods for Expression of Recombinant Proteins in Pichia pastoris, Version 1.5, 25 Invitrogen, San Diego, California). For both BPI and $L_{1-197}B_{200-456}$, protein was secreted into the medium in a small-scale batch fermentation run. 116 ng/ml were secreted for the one BPI construct assayed, and 14, 11, and 10 ng/ml were secreted for the three constructs 30 $L_{1-197}B_{200-456}$ constructs assayed. Secretion was assayed by enzyme-linked immunosorbant analysis (ELISA). majority of protein for both constructs was not secreted, as shown by Western blot analysis with a polyclonal anti-BPI antibody (INVN 1-2) (prepared by conventional

- 56 -

t chniques by inj cting rabbit with BPI) and alkaline phosphatase-conjugated goat anti-rabbit antibody. The Western blot is shown in Figure 18.

Purified BPI from Chinese Hamster ovary cells

(CHOs) was used as a positive control (lane 12). In lane

1 a sample from untransformed GS115 cells served as a
negative control. The antibodies did not recognize any
proteins from the untransformed GS115 cells. The next
three lanes (2-4) were samples from colonies transformed

with the construct for BPI and the last 6 lanes (5-10)
were samples from colonies transformed with the construct
for L₁₋₁₉₇B₂₀₀₋₄₅₆. The amount of intracellular BPI or
L₁₋₁₉₇B₂₀₀₋₄₅₆ expressed in the batch fermentation run,
based on the amount of standard BPI loaded, was roughly

15 100 μg/ml of medium for the BPI and L₁₋₁₉₇B₂₀₀₋₄₅₆
colonies.

Example 4: Protein Purification

BPI was purified from conditioned media using the following four-step purification. BPI was captured on CM Sepharose (Pharmacia LKB Biotechnology). The column was washed in 50 mM Tris pH 7.4, and protein was eluted with 50mM Tris buffer pH 7.4 + 1 M NaCl. The eluate was diluted 10% with 50mM Tris pH 8.5, run over Fast Q Sepharose, and the flow-through was collected. BPI was re-captured on CM Sepharose and again eluted as before. Buffer exchange into 10 mM Succinate + 110 mM NaCl pH 6 was performed using Sepharose CL6B (Pharmacia LKB Biotechnology). Finally, Tween 20 was added to the formulated material to a final concentration of 0.05%.

on Fast S Sepharose (Pharmacia). The column was washed with 50 mM Tris pH 7.4, and protein was eluted using 50mM Tris pH 7.4 + 1 M NaCl. The eluate was diluted 10X in 50mM Tris pH 8.5 and run over HiLoad Q Sepharose

(Pharmacia). Protein was eluted with a 0-1 M NaCl gradient in 50mM Tris pH 8.5. Appr priate fractions were pooled according to migration on SDS PAGE electrophoresis. LBP concentration was diluted to 4.0 5 mg/ml, and the pH was adjusted to 7.0 with 100 mM HCl.

 $L_{1-197(143->V)}^{B}_{200-456(N206->D)}$ was purified from cell culture medium using the same method described for LBP.

 $B_{1-199}L_{200-456}$ and $B_{(8351->A)}$ were purified using the same protocol as for BPI, except that the size exclusion 10 step was omitted.

L₁₋₅₉B₆₀₋₄₅₆, L₁₋₁₃₄B₁₃₅₋₄₅₆ and B_{CAT6} were captured on a Poros II HS cation exchange column (Perseptive Biosystems, Cambridge, MA) at pH 7.4. The column was washed with 20 mM HEPES buffer at pH 7.5, and eluted with 20 mM HEPES pH 7.5 with 1 M NaCl. The eluate was diluted 5X in 20 mM HEPES pH 7.5 and applied to a Poros HQ anion exchange column (Perseptive) with the flow-through applied directly to a POROS II HS column. The POROS II HS column was eluted with 3.3 mM acetate, 3.3mM MES and 3.3 mM HEPES, pH 6.0 with a 0-1 M NaCl gradient.

 $L_{1-359}B_{360-456}$ and $L_{(1-198)}B_{(201-456)}$ Fc were captured from conditioned medium at pH 7.4 on a Poros II HS column. The column was washed with 20mM HEPES buffer at pH 7.5, and eluted with 20mM HEPES pH 7.5 + 1 M NaCl.

25 The eluate was diluted 10X with 20mM HEPES pH 7.5, loaded on a second, smaller Poros II HS column, and eluted with 3.3 mM acetate, 3.3 mM MES and 3.3 mM HEPES, pH 6 with a 0-1 M NaCl gradient.

Example 5: BPI Activity Against N. meningitidis and N. 30 gonorrhoeae

BPI suppresses TNF release by human inflammatory cells in response to lipopolysaccharide (LPS) derived from a wide range of Gram-negative bacterial species. In order to t st the activity of BPI against Gram-negative

lipooligosaccharide (LOS) from the pathogenic bacteria
Neisseria meningitidis and N. gonorrhoeae, non-viable
bacteria were pre-treated with recombinant BPI and
incubated with human whole blood for 4 hours at 37°C.

5 Without BPI, N. meningitidis at 105 bacteria/ml
stimulated the release of 2.93 ± 0.53 ng/ml of TNF, while
N. gonorrhoeae was a more potent stimulator of TNF
release: 104 bacteria/ml induced 8.23 ± 0.32 ng/ml of
TNF. In both cases, 10 µg/ml BPI completely inhibited

10 TNF release. This indicates that BPI is able to bind and
detoxify LOS of these organisms, as well as bind LPS.
Thus, BPI can be useful as a therapeutic agent against
LOS-mediated tissue damage associated with these
pathogenic Neisseria species.

15 Example 6: biotinylated BPI Binding Competition Assays Competition assays for binding of LPS immobilized on microtiter plates was performed using a modified procedure described by Tobias et al., J. Biol. Chem. 264:10867 (1989). Briefly, Immulon 3 microtiter plates 20 (96-well, Dynatech Biotechnology Products, Chantilly, VA) were coated with 1 or 4 μg of <u>S. minnesota</u> R595 Re LPS (LIST Biological Labs, Inc., #304) in 50mM borate pH 9.5-9.8 + 20-25 mM EDTA overnight at 37°C. non-LPS coated wells were included on each plate and 25 binding to these wells was used to determine non-specific binding. Absorbance values from wells which were not pre-coated with LPS consistently gave optical density readings of less than 0.05. Plates were then washed extensively under running distilled deionized water, then 30 dried at 37°C. Assay wells were blocked for 60 minutes at 37°C with 1-2% very low endotoxin BSA (Sigma, St. Louis, MO) prepared in pyrogen-free Tris-buffered saline (50mM Tris pH 7.4 + 150 mM NaCl). The wells were emptied, and biotinylated BPI was incubated in the

presenc or absence of unlabeled BPI or recombinant pr tein f the subj ct invention dilut d in assay buffer (pyrogen-free TBS + 1 mg/ml low endotoxin BSA, and 0.05% Tween-20) was incubated in the LPS coated and uncoated 5 wells for 2-3 hours at 37°C in a total volume of 100 µl/well. After four washes in assay buffer, plates were developed with streptavidin conjugated to alkaline phosphatase (BioRad, Burlingame, California) followed by 100 µl of PNP substrate solution (Sigma) freshly prepared from two 5 mg tablets dissolved in 10 ml substrate buffer. Substrate buffer is prepared with 24.5 mg MgCl2, 48 ml diethanolamine, brought up to 400 ml, pH adjusted to 9.8 and volume brought up to 500 ml. Absorbances were read at 405 nm on a Vmax kinetic microplate reader 15 (Molecular Devices, Inc., Menlo Park, CA).

The relative LPS binding affinities of BPI, LBP and RENPs were tested in the competitive binding assay described above using 10 ng/ml biotinylatedBPI. In these experiments, BPI inhibited biotinylatedBPI binding to LPS in a concentration-dependent manner (Figure 7). Modest inhibition of biotinylatedBPI-binding was observed using NCY102 (LBP) and L_{1-197(I43->v)}B_{200-456(N206->p)}, suggesting that BPI has either a higher affinity for LPS bound to a surface or that LBP and L_{1-197(I43->v)}B_{200-456(N206->p)} bind to a different site on LPS. B₁₋₁₉₉L₂₀₀₋₄₅₆, which contains the N-terminal domain of BPI, competed with biotinylatedBPI at similar concentrations as unlabeled BPI, suggesting a similar affinity and binding site.

Competition between either $L_{1-197}B_{200-456}$ (NCY118)

30 or $L_{1-197}(143-v)^B_{200-456}(N206-v)$ (NCY103) with biotinylated BPI occurred at similar concentrations, giving overlapping curves (Figure 13A) indicating that the two amino acid differences between these two molecules $[L_{1-197}B_{200-456-v}L_{1-197}(143-v)^B_{200-456}(N206-v)$: (I43-v) and 35 (N206-vD) had no ffect on affinity for immobilized LPS.

.- 60 -

 $L_{(1-198)}B_{(201-456)}Fc$ (an IgG chimera consisting of $L_{1-197}B_{200-456}$ linked to human IgG1 Fc constant regi n of the immunoglobulin molecule) does not have an altered ability to compete with biotinylated BPI (Figure 13A).

5 L₁₋₅₉B₆₀₋₄₅₆ and L₁₋₁₃₄B₁₃₅₋₄₅₆ showed a similar affinity for LPS which affinity was very similar to that observed for BPI, suggesting that the region between amino acid residues 1-59 (or 1-134) probably plays a minimal role in LPS binding (Figure 13B). Together with data showing the B₁₋₁₉₉L₂₀₀₋₄₅₆ competes effectively with BPI (Figure 7), these results indicate that amino acid residues 134-199 are important structural components of the high-affinity

LPS-binding domain of BPI.

The importance of the region between amino acid 15 residues 134 to 197 in LPS affinity was further demonstrated by the markedly reduced affinity of BCAT9, a mutant in which all of the cationic amino acids of the BPI molecule (particularly the cationic residues of BPI amino acids 134-200) are replaced with the corresponding 20 amino acid residues found in LBP. These changes resulted in a molecule with binding affinity for LPS which was more similar to that of LBP than BPI (Figure 13C, and Figure 7). Amino acid residues 360 to 456 of BPI are apparently not involved in LPS binding as demonstrated by 25 the relative inability of $L_{1-359}B_{360-456}$ to displace biotinylated BPI from LPS (Figure 13C). The apparent binding affinity of $L_{1-359}B_{360-456}$ for LPS is similar to that of LBP and B_{CAT9}, which affinity is approximately two orders of magnitude lower than the apparent affinity of 30 BPI for LPS.

Thus, the domain of BPI which participates in binding to immobilized LPS is localized in the N-terminal half of the BPI molecule, since $B_{1-199}L_{200-456}$ has the greatest ability to displace native BPI from LPS coated onto microtiter plates. This domain of BPI has been more

. - 61 -

specifically localized to a region between amino acid residues 134-199.

Example 7: Chromogenic LAL Assay

To test the relative abilities of BPI, LBP and ENPs to neutralize LPS in vitro, these proteins were tested for inhibitory activity in the chromogenic LAL assay. Briefly, BPI and RENPs (25 μl of 0-200 μg/ml) were pre-incubated for 1 hour at 37°C with 1 EU/ml of E. coli 0111:B4 LPS, (Whitaker Biologicals, Walkersville,

- 10 MD). The mixtures were then tested for LAL activity using the chromogenic LAL assay kit (Whitaker Biologicals, Walkersville, MD). The results are shown in Figure 8 and Table 4. LPS was neutralized by the various proteins tested in the order of:
- 15 $^{\rm B}_{(8351->A)}$ $^{\geq}$ BPI $^{\rm L}_{1-197(I43->V)}^{\rm B}_{200-456(N206->D)}$ $^{\rm B}_{1-199}^{\rm L}_{200-456}$ $^{\rm LBP}.$ Several studies were carried out with different lots of each protein and the IC₅₀ values were determined. The averaged IC₅₀ values are shown in Table 4.

Table 4

LPS Inhibition in the Chromogenic LAL Assay

5	Product	IC ₅₀ (μg/ml)	No. of test
	_		
	B _(8351->A)	1.5	(n=1)
10	BPI	5.2 ± 3.3	(n=10)
	L ₁₋₁₉₇ (143->V)B ₂₀₀₋₄₅₆	S(N206->D) 28.0	± 20.0 (n=4)
	B ₁₋₁₉₉ L ₂₀₀₋₄₅₆	40.0	(n=1)
	LBP	65.0 ± 31.0	(n=4)
15	-		

15

These results demonstrate that BPI neutralizes LPS activity in the LAL assay at lower concentrations than LBP. B₁₋₁₉₉L₂₀₀₋₄₅₆, which contains the N-terminal domain of BPI, effectively competes with BPI for binding to LPS (see Figure 7) but is a relatively poor inhibitor of LPS in the LAL assay. These results indicate that the N-terminal (LPS-binding) domain of BPI alone does not account for the neutralizing activity of BPI in the LAL assay. L₁₋₁₉₇(I43->V)B₂₀₀₋₄₅₆(N206->D) was a more potent inhibitor than LBP or B₁₋₁₉₉L₂₀₀₋₄₅₆, suggesting that the C-terminal domain of BPI plays a very important role in endotoxin neutralization in the LAL assay.

Additional results of LPS neutralizing activity in the chromogenic LAL assay are shown in Table 5.

 30 $^{L_{1-197(I43->V)}}$ B $^{200-456(N206->D)}$, $^{L_{1-59}}$ B $^{60-456}$, and $^{L_{1-134}}$ B $^{135-456}$ share the C-terminal half of the BPI molecule, again indicating that this domain plays an important role in LPS-neutralizing activity. Also, these data indicate that the 199-456 region is most important in LPS

.- 63 -

neutralization since adding BPI amino acid residues betw n 136-456 or 60-456 did not improve LPS neutralizing activity. Together with the LPS binding data, these results further indicate that the C-terminal 5 half of BPI is important for neutralization, while the N-terminal sequence is more critical for LPS binding.

Table 5

LPS Inhibition in the Chromogenic LAL Assay

	The state of the s			
5	Protein		IC50	n
	BPI	Cumulative	1.58 ± 1.58	94
		Lot# 149718	1.57 ± 1.01	54
		Lot# 149719	1.69 ± 0.35	7
		Lot# 149722	1.70 ± 0.28	2
10		Lot# 149724	1.41 ± 0.45	45
		Lot# 155794	1.95 ± 0.92	2
	LBP	Cumulative	55.92 ± 30.53	8
		Lot# 151281	34.33 ± 7.45	6
		Lot# 151204	77.50 \pm 24.45	2
15	$\frac{L_{1}-197(143->v)}{22.86 \pm 16.28}$	(N206->D) 54	Cumulative	
		Lot# 151235	25.50 ± 0.71	2
		Lot# 151242	36.50 ± 0.71 36.50 ± 2.12	2
		Lot# 151274	3.46 ± 2.12 3.46 ± 2.18	2
20		Lot# 159616		38
		10CF 139010	8.83 ± 4.91	4
	B ₁₋₁₉₉ L ₂₀₀₋₄₅₆	Cumulative	24.19 + 6.42	9
	333 333	Lot# 151246	12.50 ± 0.26	3
		Lot# 152658	10.70	1
		Lot# 155737	40.18 ± 34.48	4
25	B ₁₋₁₉₉	Cumulative	5.52 ± 5.05	17
		Lot# 151285	1.12 ± 0.00	
		Lot# 155709	9.73 ± 1.18	2
		Lot# 155779	2.13 ± 0.81	3
		2007 155775	2.13 ± 0.81	2
	L ₁₋₅₉ B ₆₀₋₄₅₆	Lot# 155754	3.64 ± 1.64	5
30	L ₁₋₁₃₄ B ₁₃₅₋₄₅₆	Lot# 155756	5.02 ± 3.11	5
	L ₁₋₂₇₅ B ₂₇₈₋₄₅₆	Lot# 155791	14.00 ± 2.65	3
	L ₁₋₃₅₉ B ₃₆₀₋₄₅₆	Lot# 155733	>100	
	T_731_200-620	200# 133/33	~ 1 00	4

-	65	_

	Protein		<u>IC50</u>	n
	L ₁₋₁₉₇ B ₂₀₀₋₄₅₆	Cumulative Lot# 155758 Lot# 159619	12.75 ± 3.54 10.25 ± 30.9 15.25 ± 5.91	12 8 4
5	B _{CAT6}	Lot# 155785	1.97 ± 0.06	3
	B _{CAT9}	Lot# 155762	29.60 ± 23.23	5
	B _{CAT15}	Lot# 155788	7.87 ± 2.80	3
	$L_{(1-198)}^{L}_{(202-275)}^{L}_{(274-456)}$		Lot# 159649	
10	L ₍₁₋₁₉₈₎ B ₍₂₀₁₋₄₅₆₎ Fc	Lot# 155760	12.15 ± 6.00	4
	L ₁₋₁₉₉		9.2	1
	B ₁₋₁₉₉		10.1 \pm 0.92	5
	$^{L}_{(1-134)}^{B}_{(136-275)}^{L}_{(274-456)}^{L}_{(274-456)}$		Lot# 159643	
.15			•	

B_{CAT9}, which contains the entire BPI sequence except for nine cationic residues between positions 148 and 197 showed very poor LPS-neutralizing activity,

20 suggesting that these residues are important in LPS-neutralizing activity. Similarly, this compound was relatively ineffective at competing with native BPI for binding to LPS. These cationic residues may permit correct structural conformation of the molecule, since

25 both L_{1-197(I43->V)}B_{200-456(N206->D)} and B_{CAT9} contain the C-terminal domain of BPI, yet L_{1-197(I43->V)}B_{200-456(N206->D)} has potent neutralizing activity while B_{CAT9} does not.

Example 8: Inhibition of FITC-labeled LPS binding to human monocytes

The relative LPs-binding affinities of RENPs of the invention were investigated by examining the abilities of the RENPs to inhibit LPS binding to human

- 66 -

peripheral blood monocytes. Blood collected in acid citrate dextr se-containing VACUTAINER tubes (Becton Dickinson, Rutherford, NJ) was diluted 1:4 in Hank's balanced salt solution (HBSS) minus calcium and magnesium 5 (Gibco BRL, Grand Island, MD). Mononuclear cells were isolated using Ficol-Paque (Pharmacia Inc., Piscataway, NJ). Cells were washed three times in HBSS, then brought up to an appropriate volume of RPMI 1640 with glutamine and antibiotics to give approximately 1 X 106 cells/ml. 10 To one ml aliquots of cells, FITC-LPS was added to a final concentration of 500 ng/ml. Tubes were closed and incubated at 37°C on a rocking platform. At the end of the incubation, cells were washed twice with PBS with 0.05% Human Serum Albumin and 0.02% sodium azide. FACS 15 analysis of the cells was performed on a FACStar flow cytometer, Immunocytometry System, Becton Dickinson (Mountain View, CA). The monocyte portion of the cell population was determined by side scatter versus forward scatter gating and confirmed by staining a separate 20 aliquot of cells with phycoerythrin-conjugated anti-DR antibody (Becton Dickinson Immunocytometry Systems, Milpitas, CA). Results are reported as logarithmic scale mean fluorescence intensity.

L₁₋₁₉₇(I43->v)^B200-456(N206->D) to inhibit LPS binding to human peripheral blood monocytes, isolated human peripheral blood monocytes, isolated human peripheral blood mononuclear cells were incubated with 10% human serum containing 500 ng/ml FITC-conjugated E. coli 055:B5 LPS in the presence or absence of BPI or

30 L₁₋₁₉₇(I43->v)^B200-456(N206->D). Binding of FITC-LPS to monocytes could be inhibited by increasing concentrations of both BPI and L₁₋₁₉₇(I43->v)^B200-456(N206->D) (Figure 9). Thus L₁₋₁₉₇(I43->v)^B200-456(N206->D) has BPI-like activity, despite the fact that L₁₋₁₉₇(I43->v)^B200-456(N206->D) contains the N-terminal domain of LBP. These data, along with the

r sults of the LPS neutralization studies shown in Figure 8, suggest that the C-terminal domains of BPI and LBP, and not the N-terminal domains, determine whether the proteins inhibit or mediate LPS activation of cells.

Further studies were undertaken to determine the 5 effects of BPI, LBP, $L_{1-197(143->V)}B_{200-456(N206->D)}$ and $B_{1-199}L_{200-456}$ on FITC-labeled LPS binding to peripheral blood monocytes in the presence and absence of serum. a serum-free FITC-labeled LPS binding system where no LBP 10 is available, FITC-labeled LPS does not bind to cells. In contrast recombinant LBP facilitated LPS binding to cells at concentrations as low as 100 ng/ml. B₁₋₁₉₉L₂₀₀₋₄₅₆ also facilitated binding, although to a lesser extent. Neither BPI or $L_{1-197(143->V)}B_{200-456(N206->D)}$ 15 promoted significant binding of LPS to cells. These data indicate that the C-terminal domain of LBP is active in LPS binding to cells. The N-terminal domain of BPI may exert an inhibitory influence on LPS binding to cells mediated by the C-terminal domain of LBP because $B_{1-199}L_{200-456}$ was less active than LBP.

Normal human serum contains about 1-10 µg/ml LBP. In the presence of 10% autologous serum, BPI and L1-197(I43->v)B200-456(N206->D) potently inhibited FITC LPS binding to monocytes, with BPI showing slightly greater 25 potency. B1-199L200-456 had marginal activity, and LBP had no effect (Figure 14A). These data indicate the importance of the BPI C-terminus in this test of LPS neutralization. B1-199L200-456, which lacks the C-terminal domain of BPI, is approximately two orders of magnitude less potent at blocking LPS binding. LBP, as expected, had no effect. Thus, BPI can intervene in the sepsis cascade by preventing LPS from binding to monocytes and causing release of TNFalpha.

- 68 -

Example 9: THP-1 Cell TNF Production Assay

THP-1 cells w r obtain d from the American Tissue Culture Collection (Rockville, MD) and were maintained in REM medium containing 10% fetal bovine serum, 2 mM 5 L-glutamine, 100 units penicillin and 100 μ g/ml streptomycin. Cells were passed at 2 x 105 cells/ml every 3 days. Responsiveness of THP-1 cells to LPS was induced by culturing the cells for 48 hours in REM medium containing 10% fetal calf serum, 2 mM L-glutamine, 100 10 units penicillin, 100 μ g/ml of streptomycin and 100 nM PMA at 37°C in a humidified atmosphere with 5% CO2. Cells were cultured in 96-well flat-bottomed tissue culture plates at 1-2 x 105 cells per well in a final volume of 200 μ l. After 48 hours, adherent cells 15 were washed three times with 200 μ l of medium without serum. To 180 μ l of medium without serum but with 0.5% HSA, LPS (10 μ l at 200 ng/ml) and/or BPI, LBP or other RENPs were added (10 μ l at 0-2 mg/ml) and the cells were cultured for an additional 4 hours. After 4 hours, 20 supernatants were transferred to a U-bottomed 96 well plate and the plate was centrifuged (500 x g, 12 min) to pellet any cell debris. Supernatants were then stored in a second plate at -20°C until assayed for TNF by ELISA.

To further identify the regions of BPI which

25 contribute to LPS-neutralizing activity, and the domains
of LBP which are responsible for transducing the LPS
signal to cells, the abilities of RENPs to replace LBP
were compared under serum-free conditions. In these
experiments, cells of the promonocytic cell line THP-1

30 were induced to respond to LPS by culturing for 48 hours
with phorbol ester. After induction, cells were
stimulated with 19 ng/ml of LPS in the presence or
absence of the recombinant protein. In this system, TNF
release requires a source of LBP. Data from these

35 experiments (Figure 15) show that only LBP and

- 69 -

L₁₋₃₅₉B₃₆₀₋₄₅₆ stimulated TNF release. Thus the domain of LBP responsible for facilitating LPS-induced TNF release is within amino acid residues 199-359. Interestingly, B₁₋₁₉₉L₂₀₀₋₄₅₆ did not mediate TNF release in a serum-free system. This may indicate that the N-terminal domain of BPI binds too tightly to LPS to allow transfer of LPS to CD14 on the macrophage surface. Figure 16 shows an additional comparison of TNF production. Because L₍₁₋₁₉₈₎B₍₂₀₂₋₂₇₅₎L₍₂₇₄₋₄₅₆₎ includes LBP domain 274-456 and has activity, the active domain may comprise only residues 274-359.

Example 10: LPS-Induced TNF Release In Whole Blood

Peripheral blood from normal human volunteers was collected in heparin-containing VACUTAINER tubes (Becton Dickinson). To one milliliter of whole blood, BPI, a protein of the subject invention, or buffer control was added, followed by 1 ng/ml E. coli 055:B5 refined standard endotoxin (RSE) (Whitaker Bioproducts). Samples were incubated in closed microtubes at 37°C for 4 hours on a rocking platform. At the end of the incubation, samples were centrifuged for 5 minutes at 500 x g at 4°C, the plasma collected and frozen on dry ice until assayed for the presence of cytokines. TNF levels were determined by ELISA using human recombinant TNF (Genentech Inc., South San Francisco, CA) as a standard.

In later studies it was determined that BPI activity in whole blood is inhibited by heparin, and the anticoagulant was changed to citrate. In these experiments, to 120 µl of citrated whole blood, 20 µl of 30 BPI or an RENP (at 0-1 mg/ml) or buffer control, 20 µl of 100 ng/ml of E. coli 055:B5 LPS was added to stimulate cells in whole blood samples. These experiments were performed in polypropylene microtiter plates (Costar, Cambridge, MA). After the 37°C incubation step, the

plates were c ntrifuged 15 min at 500 \times g at 4°C and the plasma removed f r assaying.

To test the effects of BPI, LBP, and RENPs on LPS activation of TNF production in whole blood, BPI, LBP,

- 5 L_{1-197(143->v)}B_{200-456(w206->D)}, or B₁₋₁₉₉L₂₀₀₋₄₅₆ was mixed with heparinized blood, and LPS was added to the resulting mixture. The blood was incubated for four hours at 37°C, and TNF in the plasma was measured as described, supra. Results are shown in Figure 10.
- 10 $^{L_{1-197(I43->V)}B_{200-456(N206->D)}}$ was the most potent at blocking TNF release, followed by BPI as the next most potent blocker. $B_{1-199}L_{200-456}$ and LBP had essentially no effect. Thus, in whole blood, $L_{1-197(I43->V)}B_{200-456(N206->D)}$ proved to be the most potent inhibitor of LPS-mediated cytokine stimulation.

When experiments were performed in citrated rather than heparinized whole blood, endotoxin-neutralizing activity of BPI and $L_{1-197(I43->V)}^{B}_{200-456(N206->D)}$ were equivalent (Table 6). In experiments in which

- 20 recombinant proteins were preincubated with endotoxin before addition to whole blood, the activities of these compounds fell roughly into two groups. BPI,
 - $L_{1-197(143->V)}^{B}_{200-456(N206->D)}$, B_{1-199} , B_{CAT6} , B_{CAT15} , $L_{1-59}^{B}_{60-456}$, $L_{1-134}^{B}_{135-456}$, and $L_{1-197}^{B}_{200-456}$ possess
- LPS-neutralizing activity, while LBP, $B_{1-199}L_{200-456}$, L_{1-199} , $L_{1-359}B_{360-456}$ and B_{CAT9} were relatively inactive. Results with $L_{1-275}B_{278-456}$, B_{CAT9} , and $L_{(1-198)}B_{(201-456)}F_{CM}$ were equivocal. When compounds were added to the blood samples immediately prior to LPS, the IC50 values were
- 30 higher, but the same group of proteins showed activity. These data further indicate the role of the C-terminal region of BPI, demarcated by amino acid residues 200-359, in LPS neutralization in a physiological environment such as whole blood. Because L_{1-199} is not a potent
- 35 endotoxin-neutralizing protein (see Tables 9 and 11), it

- 71 -

can be concluded that th C-terminal domain of BPI must significantly c ntribute t the endotoxin-neutralizing activity of $L_{1-197(143->V)}B_{200-456(W206->D)}$ and $L_{1-197}B_{200-456}$. All compounds which contain this sequence (201-359) are active except B_{CAT9} , which was also inactive in other assays possibly because the cationic amino acid residues which were replaced may be important in configuring the molecule. These data indicate that

L₁₋₁₉₇(143->v)^B200-456(W206->D) is equivalent to L₁₋₁₉₇B₂₀₀₋₄₅₆
10 in activity, thus implying that the amino acid
differences between these two proteins have no affect
upon LPS binding affinity.

Table 6

LPS Inhibition in Human Whole Blood

5	Protein Pre- incubated	1C50 (μg/ml)	n	Not Pre- incubated	IC50 (μg/ml)	n
	L ₁₋₁₃₄ B ₁₃₅₋₄₅₆	0.15 ± 0.12	3	BPI	2.60 ± 1.52	5
	L _{1~197} B _{200~456}	2.90 ± 3.59	1 2	L ₁₋₁₃₄ B ₁₃₅₋₄₅₆	3.7 ± 1.60	2
	L ₁₋₅₉ B ₆₀₋₄₅₆	0.28 ± 0.25	3	L ₁₋₁₉₉ \7(I43->V) ^B 200-456 (N206->D)	7.13 ± 5.92	4
10	L ₁₋₁₉₇ (I43->V) ^B 200-456(N208->D)	0.16 ± 0.11	1 7	L ₁₋₅₉ B ₆₀₋₄₅₆	15 ± 18.58	2
	BPI	0.43 ± 0.49	1 3	L ₁₋₁₉₇ B ₂₀₀₋₄₅₆	26.5 ± 0.71	2
	L ₍₁₋₁₉₈₎ B ₍₂₀₁₋₄₅₈₎ PC	18.00 ± 27.73	3	L ₁₋₃₅₉ B ₃₆₀₋₄₆₆	>100	1
	B ₁₋₁₉₉ L ₂₀₀₋₄₅₆	>100	3	B _{CAT9}	>100	2
	L ₁₋₃₅₉ B ₃₆₀₋₄₅₆	>100	3	L ₍₁₋₁₉₈₎ B ₍₂₀₁₋₄₅₆₎ Fc	>100	2
15	B _{CAT9}	11.50 ± 3.54	2	B ₁₋₁₉₉ L ₂₀₀₋₄₅₆	ND	
	B ₁₋₁₉₉	0.73 ± 0.48	6	B ₁₋₁₉₉	4.0	1
	L ₁₋₁₉₉	>100	2	L ₁₋₁₉₉	>100	1
	B _{CAT15}	0.21 ± 0.26	3			
	B _{CAT6}	0.27 ± 0.25	2			
20	L ₍₁₋₁₃₄₎ B ₍₁₃₆₋₂₇₅₎ L ₍₂₇₄₋₄₅₆₎	2.0	1			
	L ₍₁₋₁₉₈₎ B ₍₂₀₂₋₂₇₅₎ L ₍₂₇₄₋₄₅₆₎	5.27 ± 5.83	3			
	L ₁₋₂₇₆ B ₂₇₈₋₄₅₆	38.10 ± 53.64	3			

^{25 *}Two additional values for B_{CAT9} were >100.

.- 73 -

Example 11: Mouse Serum Half-Life Assay

CD-1 mice weighing approximately 20 grams were injected with 5 mg/kg body weight BPI, LBP, or RENPS (1 mg/ml) at time zero. In heparinized (or later 5 EDTA-containing) tubes, blood was collected from the retroorbital plexus from three animals for each time point tested. A typical blood collection schedule was 5, 10, 15, 30, 45, 60, 90, 120, 240, and 360 minutes. The EDTA anticoagulated blood was centrifuged for about 10 min at 1000 x g and the supernatant plasma removed and stored frozen on dry ice until tested. Levels of BPI, LBP, or RENP in the plasma samples were determined by ELISA using the appropriate protein as the standard.

A potent anti-endotoxin therapeutic should not
only neutralize endotoxin, but should also have the
capacity to clear endotoxin from the circulation. The
circulating levels of radioactively labeled ¹²⁵I-BPI were
measured in the mouse in the presence and absence of
endotoxin (Table 7). In the absence of endotoxin, the
elimination (alpha) phase for ¹²⁵I-BPI was less than two
minutes. In the presence of LPS, the alpha phase was
extended to 6.2 minutes. ¹²⁵I-LPS alone has a single
phase distribution (beta) with a half-life of about 101
minutes. When ¹²⁵I-LPS and unlabeled BPI were
administered, a 6.2 minute elimination (alpha) phase was
observed, indicating that elimination was remarkably
facilitated by BPI.

Table 7
Serum Half-Life of BPI and LPS in the Mouse

5	Test Article	t _{1/2} alpha	t _{1/2} beta	
	¹²⁵ I-BPI ¹²⁵ I-BPI + LPS	1.6 6.3	103.0 72.0	
	125 _{I-LPS} 125 _{I-LPS} + BPI	6.2	101.0 114.0	
10				

In order to determine whether the very short circulating half-life of BPI could be extended by molecular engineering, the circulating half-lives of BPI,

- 15 LBP, B₁₋₁₉₉L₂₀₀₋₄₅₆ and L_{1-197(I43->V)}B_{200-456(N206->D)} were compared (Figure 11). Using both labeled and unlabeled material, it was observed that the circulating half-life of BPI in the mouse is remarkably short. This may be a result of the highly cationic nature of BPI having a
- 20 predicted pI of 10.6. LBP, normally present in the circulation at concentrations of 10 μg/ml, has a predicted pI of about 6.8. As expected,
 L_{1-197(I43->V)}^B_{200-456(N206->D)} (LBP-BPI chimera lacking BPI cationic residues) has a markedly longer circulating
- 25 half-life than $B_{1-199}L_{200-456}$ (BPI-LBP chimera having BPI cationic residues). Figure 11 shows that $L_{1-197(143->v)}B_{200-456(N206->D)}$ indeed has a longer half-life than BPI. $B_{1-199}L_{200-456}$, with the N-terminal domain of BPI, had an even shorter half-life than that of BPI.
- 30 Thus, the N-terminal domain of BPI appears to play a major role in its short circulating half-life.

Further pharmacokinetic studies were performed in which recombinant proteins of the subject invention were administered to CD-1 mice at a 5 mg/kg bolus dose.

35 Results of these experiments are shown in Figures

17A-17H. At 5 mg/kg, the circulating half life of B₁₋₁₉₉L₂₀₀₋₄₅₆ was similar to that of BPI. $L_{1-197(143->V)}B_{200-456(N206->D)}$ and $L_{1-197}B_{200-456}$ had overlapping elimination curves and again indicating that 5 these two molecules are equivalent with respect to their biological activities. $L_{1-197(143-v)}^{B_{200-456(N206-v)}}$ and B_{1-199} persisted in the circulation significantly longer than BPI or $B_{1-199}L_{200-456}$, but not as long as the serum protein LBP. Comparison of the elimination curves of 10 $L_{1-59}B_{60-456}$, $L_{1-134}B_{135-456}$ and B_{CAT9} revealed that the N-terminus of LBP plays a role in extending circulating half-life. L₁₋₅₉B₆₀₋₄₅₆ circulates slightly longer than BPI, and contains the least LBP sequence of any of the recombinant proteins tested (amino acid residues 1-59). 15 $L_{1-134}B_{135-456}$ was cleared somewhat more slowly, indicating that LBP amino acid residues 60-134 of LBP impart a longer circulating half-life. In contrast, the cationic residues of BPI between 134-199 shorten the half-life, since in B_{CAT9} , where the cationic residues in this region 20 were replaced with the corresponding residues of LBP, the half-life was similar to that of $L_{1-134}B_{135-456}$. more LBP residues in the N-terminal domain further extends the half life. If amino acid residues 199-359 of LBP are added $(L_{1-359}B_{360-456})$, the half-life is longer, 25 but not quite as long as that of LBP. Likewise $L_{(1-198)}^{B}_{(202-275)}^{L}_{(274-456)}$ (with LBP domain 1-198 and 274-456) has a relatively long t1/2. These results indicate that the more "LBP-like" the molecule is, the longer it circulates. In addition, combining an Ig 30 fragment Fc with $L_{1-197(143->V)}^{B_{200-456(N206->D)}}$ gives the

Example 13: Mouse Endotoxin Challenge Assay

longest half life.

Female CD-1 mice were injected in the lateral tail vein with a LD_{100} dose (25-35 mg/kg) of <u>Salmonella abortus</u>

equi endotoxin, which was followed by an injection f
BPI, RENP, or vehicle c ntrol into th opposite lateral
tail vein at the indicated time. Protein injection
concentrations varied and provided doses of 0.5, 1, and
5 mg/kg. Use of vehicle control illustrated the
lethality of the endotoxin challenge in the test animal.
Animals were observed for mortality at 24, 28, and 72
hours. Preliminary studies showed that mortality does
not change from day three to day seven or beyond.

The efficacies of BPI, LBP,

 $L_{1-197(143->V)}^{B_{200-456(N206->D)}}$, $B_{1-199}^{L_{200-456}}$ and $B_{(S351->A)}$ against lethal endotoxin challenge in mice were compared (Tables 8-10). The efficacies of

- $L_{1-197(143-v)}^{B}_{200-456(N206-v)}$, $L_{1-197}^{B}_{200-456}$, $L_{1-59}^{B}_{60-456}$, $L_{1-134}^{B}_{135-456}$, $L_{(1-198)}^{B}_{(201-456)}^{F}_{C}$, $L_{1-275}^{B}_{278-456}$, $L_{1-359}^{B}_{360-456}$, L_{CAT9} , L_{CAT6} , and L_{CAT15} against lethal endotoxin challenge in mice were also compared (Table 11). When each protein was given within two minutes after lethal endotoxin challenge, BPI,
- L_{1-197(I43->V)}B_{200-456(N206->D)} and B_(8351->A) had similar potency, whereas LBP and B₁₋₁₉₉L₂₀₀₋₄₅₆ showed minimal protection. The marginal protective effects of LBP and B₁₋₁₉₉L₂₀₀₋₄₅₆ since these agents do not block the inflammatory signal of LPS acting on human cells *in vitro* (Figure 10).

5

Table 8

Mouse Endotoxin Challenge Comparison of BPI, LBP (NCY102), and $L_{1-197}(143->v)^{\underline{B}_{200-456}(N206->D)}$ (NCY103)

	Drug	Dose	<pre>\$ Survival (n=10)</pre>
	Control	0 mg/kg	0%
10	BPI	5 mg/kg 1 mg/kg	60% 40%
	LBP	5 mg/kg 1 mg/kg	30% 20%
15	L ₁₋₁₉₇ (143->V) ^B 200-456(8	1206->b) 1 mg/kg	5 mg/kg 60% 50%

Table 9

Mouse Endotoxin Challenge

Comparison of BPI, L₁₋₁₉₇(143->v)^B200-456(N206->D) and
B(8351->A)

	Drug	<u>pose</u>	* Survival (n=10	77
	Control	0 mg/kg	0%	
	BPI	5 mg/kg	80%	
25	L ₁₋₁₉₇ (143->v) ^B 200-456(1	W206->D)	5 mg/kg	100%
	B _(8351->A)	5 mg/kg	90%	

5

20

Table 10

Mouse Endotoxin Challenge Comparison of BPI and B₁₋₁₉₉L₂₀₀₋₄₅₆ (NCY104)

	Drug	Dose	<pre>% Survival (n=10)</pre>
	Control	0 mg/kg	40%
10	BPI	10 mg/kg 2 mg/kg 0.4 mg/kg	100ዩ 100ዩ 70ዩ
	B ₁₋₁₉₉ L ₂₀₀₋₄₅₆	10 mg/kg 2 mg/kg 0.2 mg/kg	60% 60% 50%
15	-		

Table 11

<u>Survival in CD-1 Mice Following</u> <u>Lethal Endotoxin Challenge</u>

Panel A

		Survivors/n	<pre>\$ Survival</pre>	p (vs.control)
	BPI	40/50	80.00	< 0.001
25	L ₁₋₁₉₇ (I43->V) ^B 200-45	66 (N206->D)	17/20	85.00<
	L ₁₋₁₉₇ B ₂₀₀₋₄₅₆	16/20	80.00	< 0.001
	L ₁₋₅₉ B ₆₀₋₄₅₆	13/20	65.00	< 0.001
	L ₁₋₁₃₄ B ₁₃₅₋₄₅₆	13/20	65.00	< 0.001
30	(T_TAD) [(TOT-#29)	5/10	50.00	0.002
	L ₁₋₃₅₉ B ₃₆₀₋₄₅₆	2/10	20.00	0.149
	B _{CAT6}	9/10	90.00	< 0.001
	BCAT9	1/10	10.00	0.442
	L1-275B278-456	0/10	0	
35	BCAT15	6/10	60.0	< 0.05
	Control	1/30	3 30	

. - 79 -

Panel B

		Dose _mg/kg	Survivors	s { Survival	p (vs. control) *
	BPI				
5		5	13	65	< 0.001
		1	9	45	0.001
		0.5	6	30	0.02
	L ₁₋₁₉₇₍₁₄₃	->v) ^B 200-	·456 (พิ <u>२</u> 06–>ก))	
10		5		90	<0.001
10		1	12	60	<0.001
		0.5	9	45	0.001
	B ₁₋₁₉₉				
		5 .	3	15	NS
		1	0	0	NS
15		0.5	1	5	NS

^{*} Fisher's Exact Test

 $L_{1-197(143-v)}^{B}_{200-456(N206-v)}$, or perhaps the increased ability of $L_{1-197(143-v)}^{B}_{200-456(N206-v)}$ to inhibit endotoxin in whole blood, has a dramatic effect on $L_{1-197(143-v)}^{B}_{200-456(N206-v)}$ efficacy in vivo.

Further experiments were performed to assess the LPS-neutralizing activities of recombinant proteins of the subject invention in vivo. In these experiments, a lethal LPS challenge was administered at time zero, followed immediately by a 5 mg/kg bolus injection of recombinant protein.

L₁₋₁₉₇(143->v)^B200-456(N206->D) was markedly more effective than BPI when given more than an hour before or after LPS (Figure 12). These results indicate that the longer circulating half-life of

- 80 -

Example 12: BPI Reduction of LPS-Induced Cytokine Function and Mortality in Rats

The potential effect of BPI against LPS related cytokine formation and mortality was investigated in rats 5 with either (a) hemorrhagic shock (bled to lower pressure to 30-35 mmHg mean arterial pressure for 90 minutes, followed by reinfusion of shed blood and an equal volume of Ringer's over 30 minutes), or (b) endotoxic shock (caused by 100 µg LPS and 500 mg D-galactosamine/kg).

10 Similarly, recombinant BPI binds LPS and inhibits TNF formation in vitro. Treatment comprised 5 mg BPI/kg i.v. for the BPI group, or 1 ml saline i.v. for the control group.

The results of the investigation of BPI efficacy 15 in rats with either (a) hemorrhagic shock or (b) endotoxic shock show that (a) in rats with hemorrhagic shock, the mortality was decreased from 5/10 (50% control group) to 2/10 (20% BPI group) at 48 hours; (b) in rats with endotoxic shock, the 5-day mortality was 20 significantly reduced (p = 0.055) by BPI treatment to 43%, as compared to 83% in the control group. Plasma LPS levels were at least partially neutralized at two hours (5.9 \pm 4.1 vs 10.8 \pm 4.1 ng/ml). Cytokine formation was concomitantly reduced in the BPI group as measured by 25 plasma TNF levels at two hours (3.9 \pm 2.9 vs 10.3 \pm 6.3 ng/ml). Liver transaminases (GOT and GPT, whose elevation indicates hepatic dysfunction) and bilirubin still increased at eight hours; however, the increase was less with BPI. These data demonstrate that BPI has 30 utility as a therapeutic agent against endotoxin-related disorders in hemorrhagic as well as endotoxic shock.

Example 14: Protection against LPS challenge by intrapulmonary delivery of RENPS

Anesthetized male CD-1 mice were treated intra-nasally with 1 or 10 μg of either BPI or

- 5 L₁₋₁₉₇(143->v)^B200-456(N206->D) in 50 μl. Control animals received 50 μl of saline for injection. After 20 minutes, animals were re-anesthetized, and challenged with 10 ng of E. coli 055:B5 LPS. One hour after endotoxin challenge, mice were re-anesthetized, and
- 10 0.7 ml of saline containing 1% human serum albumin was added to the lungs via the trachea. The lungs were gently kneaded. A 0.5 ml volume of BAL (bronchoalveolar lavage) fluid was aspirated, cells were pelleted by centrifugation, and the BAL sample was sorted at -70°C.
- 15 The TNF-alpha level in the BAL fluid was determined by ELISA (results shown in Figure 19).

Figure 19 shows that endotoxin-neutralizing proteins such as BPI and $L_{1-197(143->V)}^{B}_{200-456(N206->D)}$ (NCY103) can also neutralize endotoxin-mediated TNF

- 20 release in the lung. These results indicate that these proteins are effective when delivered directly into the lung and thus may be useful for treatment of pneumonias and other endotoxin-related disorders of the lung, such as ARDS.
- 25 Example 15. Construction of L₁₋₁₉₇B₂₀₀₋₄₅₆

cDNA encoding $L_{1-197}B_{200-456}$ was constructed by creating a unique ClaI site at the junction between the nucleotide sequence coding for $Ile_{197}-Asp_{198}$ residues (ATA-GAT -> ATC-GAT). For $L_{1-197}B_{200-456}$, a 0.7 kb

30 NheI/ClaI DNA fragment (encoding amino acids 1-197) derived from the 5' sequence of LBP and a 0.8 kb ClaI/XhoI fragment (encoding amino acids 200-456) derived from the 3' sequence of BPI were generated by PCR. The chimeric cDNAs were spliced together by cloning the

fragments into pSE, a mammalian vector. The cDNAs for BPI, LBP and L₁₋₁₉₇B₂₀₀₋₄₅₆ w re transfected into Chinese hamster ovary cells (strain DUXB11) using lipofectin. The resulting transformed cells were selected, and expression was amplified with methotrexate. Cell culture supernatants were screened for reactivity by ELISA. Recombinant BPI, LBP, and L₁₋₁₉₇B₂₀₀₋₄₅₆ were purified as described above.

Example 16. Pharmokinetics of L₁₋₁₉₇B₂₀₀₋₄₅₆

Data for pharmacokinetic analysis were collected from healthy CD-1 mice given a single bolus injection (5 mg/kg) of recombinant protein at time=0. Blood was collected from three mice for each collection time point by retroorbital puncture at timepoints over three hours.

Blood samples anticoagulated in EDTA were assayed by a double antibody sandwich ELISA for the presence of BPI, LBP or $L_{1-197}B_{200-456}$). Pharmacokinetic analysis was performed using a non-compartmental analysis (PharmK pharmacokinetic software, SoftRes, Inc.).

Comparison of BPI and LBP shows that BPI was cleared rapidly with a clearance rate of 13.0 ml/minute (Table 12). LBP had the longest half life, with a clearance rate of 0.042 ml/min. Compared to BPI, LBP was cleared 310 times more slowly. L₁₋₁₉₇B₂₀₀₋₄₅₆ had an intermediate half life (Clearance rate = 0.175 ml/min), being cleared 74 times more slowly than BPI.

Table 12: Clearance rate of L1-197B200-456

	CL (ml/min)	(vs. BPI)
BPI	13.000	-
LBP	0.042	(310 fold)
L ₁₋₁₉₇ B ₂₀₀₋₄₅₆	0.175	(74 fold)

- 83 -

Example 17. LPS protection by L1-197B200-456

Female CD-1 mice (n=10) were inj ct d in the lateral tail vein with 35 mg/kg S. abortus equi LPS (Sigma, St. Louis, MO) at time=0. Recombinant protein 5 (5 mg/kg) was then administered intravenously into the opposite lateral tail vein immediately following (t=0) endotoxin challenge. Survival was monitored at 24, 48 and 72 hours post-challenge. Control animals received 0.1 ml saline instead of recombinant protein. The p values were determined by Fisher's exact test.

The results are shown in Figure 20. BPI and L₁₋₁₉₇B₂₀₀₋₄₅₆ provided 90% to 100% survival, respectively, at the 72 hour end point. No further mortality was noted at seven days post-challenge. The untreated control group had a survival rate of 20%. The survival rates of the treated groups were statistically significant compared to the control group (p<.001 for the L₁₋₁₉₇B₂₀₀₋₄₅₆ group and p=.003 for the BPI group determined by Fisher's exact test). These results indicate that L₁₋₁₉₇B₂₀₀₋₄₅₆ is as effective as BPI in this endotoxin challenge model in vivo.

Example 18. Protection Against Endotoxin Challenge in Mice

The ability of the recombinant,

25 endotoxin-neutralizing proteins $B_{(1-41)}L_{(1-199)}B_{(1-456)}$, $L_{(1-164)}B_{(200-456)}$, $B_{(1-175)}B_{(200-456)}$, $B_{(1-236)}$, and $B_{(1-190)}$ to protect mice against endotoxin challenge was carried out as described in Example 17 above. Protection by these proteins was compared to the protection provided by BPI or saline. The results of these studies are shown in Table 13.

- 84 -

TABLE 13

					N			Surviv (hou		10	
		Compound	Lot #	0	12	18	24	36	48	60	7 2
	Group 1	native BPI	149724	10	10	10	10	10	9	9	9
5	Group 2	B ₍₁₋₄₁₎ L ₍₁₋₁₉₉₎ B ₍₁₋₄₅₆₎	162303	10	10	10	10	9	9	8	8
	Group 3	L ₍₁₋₁₆₄₎ B ₍₂₀₀₋₄₅₆₎	164325	10	10	9	9	8	8	7	7
	Group 4	L ₍₁₋₁₇₅₎ B ₍₂₀₀₋₄₅₆₎	164326	10	10	10	10	10	10	10	1 0
10	Group 5	B ₍₁₋₂₃₆₎	159695	8	7	5	4	ı	0	0	0
	Group 6	B ₍₁₋₁₉₀₎	159699	10	9	8	6	6	6	5	5
15	Group 7	Saline		10	8	7	6	4	3	3	3

Each animal received 35 mg/kg LPS in 0.1 ml, followed immediately by 5 mg/kg of the indicated compound in 0.1 ml. Survival was monitored at each time point indicated.

20 Example 19: Detection of a Gram-negative infection in a patient

A blood sample of about 1 ml to 5 ml is drawn from a patient suspected of having a Gram-negative infection. The blood sample is treated with citrate anti-coagulant and plasma is separated from the blood cells by centrifugation. The plasma is then diluted in a series of 10-fold dilutions in assay buffer (pyrogen-free TBS + 1 mg/ml low endotoxin BSA, and 0.05% Tween-20). The diluted plasma samples are then mixed with a known amount of biotinylated RENP. A series of control samples containing known amounts of biotinylated RENP in assay

buffer is includ d in the assay as quantitativ and negative contr ls.

The test and control samples are then applied to the wells of a microtiter plate having bound LPS. The

5 LPS-bound microtiter wells are prepared by incubation with 1 or 4 μg of S. minnesota R595 Re LPS (LIST Biological Labs, Inc., \$304) in 50 mM borate pH 9.5-9.8 + 20-25 mM EDTA overnight at 37°C. Blank, non-LPS coated wells are included on each plate as controls for 10 non-specific binding. The plates are then washed extensively under running distilled deionized water, then dried at 37°C. The assay wells are subsequently blocked for 60 minutes at 37°C with 1-2% very low endotoxin BSA (Sigma, St. Louis, MO) prepared in pyrogen-free

15 Tris-buffered saline (50 mM Tris pH 7.4 + 150 mM NaCl).

The test and control samples are incubated for a time sufficient for binding of the RENP in the samples to the LPS bound to the microtiter wells, generally about 2-3 hours at 37°C in a total volume of 100 μ l/well.

- 20 After incubation, the wells are washed four times with assay buffer, and the plates are developed with streptavidin conjugated to alkaline phosphatase followed by 100 μ l of PNP substrate solution freshly prepared from two 5 mg tablets dissolved in 10 ml substrate buffer.
- 25 Substrate buffer is prepared with 24.5 mg MgCl₂, 48 ml diethanolamine, brought up to 400 ml, pH adjusted to 9.8 and volume brought up to 500 ml. Absorbances are read at 405 nm on a microplate reader.

If the level of biotinylated RENP bound to the
30 wells of the test sample is significantly less than the
level of biotinylated RENP bound to the negative control
sample, then the patient has endotoxin circulating in the
bloodstream which is generally associated with a
Gram-negative infection. Moreover, the level of RENP
35 binding in the test sample is compared to the levels of

.- 86 -

RENP binding in the quantitative controls, each of which ar r presentativ of varying degrees f severity of Gram-negative infection in a patient. The level of binding of the test sample is thus compared to the levels of binding of the quantitative samples to determine a degree of severity of infection.

Example 20: Detection of a Gram-negative infection in vivo

RENP is detectably labeled with ¹²⁵I using methods

10 well known in the art. Approximately 100 μg of an

125I-labeled RENP is injected intravenously into a patient suspected of having a Gram-negative infection in an organ, e.g., the liver. After allowing a time sufficient for circulation of the ¹²⁵I-labeled RENP to the suspected

15 site of infection, the abdomen of the patient is fluoroscoped or X-rayed 2 to 3 times so as to include various perspectives. The X-ray is then examined to identify sites of binding of the RENP by virtue of an abnormally darkened section on the X-ray. Upon

20 identification of the site of infection, the clinician designs an appropriate therapeutic regimen.

Following procedures similar to those described above, other recombinant, LPS-binding proteins can be produced and used in diagnostic methods and methods of treatment according to the invention.

The invention now being fully described, it will be apparent to one of ordinary skill in the art that many changes and modifications can be made thereto without departing from the spirit or scope of the appended 30 claims.

CLAIMS - 87 -

1. A method of detecting a site of Gram-negativ bacterial infection in a subject, said method comprising the steps of:

injecting into the patient's circulatory system an injectable formulation comprising an effective amount of a recombinant endotoxin-neutralizing polypeptide attached to a detectable label, wherein the polypeptide is characterized by (i) selective and specific binding to lipopolysaccharide and (ii) endotoxin-neutralizing activity, with the proviso that the amino acid sequence of the polypeptide is not identical to the amino acid sequences of BPI or LBP;

allowing the detectably labeled polypeptide sufficient time to circulate in the subject and bind to lipopolysaccharide in the patient; and

detecting a site of label binding in the patient, thereby detecting a site of Gram-negative bacterial infection.

- The method of claim 1, wherein the
 polypeptide is covalently bound to a molecule which enhances the half-life of the polypeptide.
 - 3. The method of claim 1, wherein the polypeptide contains an LPS binding domain of BPI, LBP, a BPI variant, or an LBP variant.
- 25 4. The method of claim 1, wherein the detectable label is a radionucleotide.

5. A m thod of detecting a Gram-negative bacterial infecti n in a subject, said method comprising the steps of:

obtaining a sample from a patient suspected of baving a Gram-negative bacterial infection;

contacting said sample with a detectably labeled recombinant endotoxin-neutralizing polypeptide for a time sufficient for binding of the polypeptide to lipopolysaccharide in the sample; and

detecting formation of lipopolysaccharide-polypeptide complexes by detection of a detectable label bound to the polypeptide;

10

wherein detection of a level of detectable label in said sample significantly greater than a level of detectable label in a negative control sample is indicative of a Gram-negative bacterial infection in the subject.

- 6. The method of claim 5, wherein the polypeptide contains an LPS binding domain of BPI, LBP, a 20 BPI variant, or an LBP variant.
 - 7. The method of claim 5, wherein said detection is quantitative.
- 8. The method of claim 7, wherein said quantitative detection is correlated with an25 Gram-negative bacterial infection load.

- 89 -

- 9. A detectably labeled recombinant endotoxin-neutralizing p lypeptid characterized by (i) selective and specific binding to lipopolysaccharide and (ii) endotoxin-neutralizing activity, with the proviso that the amino acid sequence of the polypeptide is not identical to the amino acid sequences of BPI or LBP.
 - 10. A polypeptide according to claim 9, wherein the polypeptide contains an LPS binding domain of BPI, LBP, a BPI variant, or an LBP variant.
- 11. A detectably labeled polypeptide according to claim 9, wherein the polypeptide comprises a molecule which enhances the half-life of said polypeptide and is covalently bound to the polypeptide.
- 12. A detectably labeled polypeptide according to claim 11, wherein said molecule is an immunoglobulin fragment, a half-life enhancing porion of LBP, a half-life enhancing portion of an LBP variant, or polyethylene glycol.

- 90 -

- 14. A recombinant endotoxin-neutralizing polyp ptide characterized by (i) selective and specific binding to lipopolysaccharide and
- (ii) endotoxin-neutralizing activity, with the proviso 5 that the amino acid sequence of the polypeptide is not identical to the amino acid sequences of BPI or LBP.
- 15. A recombinant endotoxin-neutralizing polypeptide according to claim 14, wherein said polypeptide is of the formula L₁₋₁₉₇B₂₀₀₋₄₅₆ or a
 10 corresponding protein which (a) functions to bind lipopolysaccharide and (b) neutralizes endotoxin.
- 16. A recombinant endotoxin-neutralizing
 polypeptide according to claim 14, wherein the
 polypeptide is BPI(S351->X), wherein X is any amino acid
 .15 other than serine.
 - 17. A recombinant endotoxin-neutralizing polypeptide according to claim 16, wherein X is alanine.
- 18. A recombinant endotoxin-neutralizing polypeptide according to claim 14, wherein the
 20 polypeptide contains the amino acid sequence of BPI having a cationic amino acid substituted with a neutral or anionic residue.
- 19. A recombinant endotoxin-neutralizing polypeptide according to claim 18, wherein the cationic 25 amino acid is at BPI amino acid residue positions 27, 30, 33, 42, 44, 48, 59, 77, 86, 90, 96, 118, 127, 148, 150, 160, 161, 167, 169, 177, 185, or 198.

- 91 -

- 20. A recombinant endotoxin-neutralizing polypeptid according to claim 19, wher in the polypeptide contains neutral or anionic residues at BPI amino acid residue positions 27, 30, 33, 42, 44, 48, and 5 59.
- 21. A recombinant endotoxin-neutralizing polypeptide according to claim 19, wherein the polypeptide contains neutral or anionic residues at BPI amino acid residue positions 77, 86, 90, 96, 118, and 10 127.
- 22. A recombinant endotoxin-neutralizing polypeptide according to claim 19, wherein the polypeptide contains neutral or anionic residues at BPI amino acid residue positions 148, 150, 160, 161, 167, 15 169, 177, 185, and 198.
- 23. A recombinant endotoxin-neutralizing polypeptide according to claim 18, wherein the polypeptide contains neutral or anionic residues at BPI amino acid residue positions 27, 30, 33, 42, 44, 48, 59, 77, 86, 90, 96, 118, 127, 148, 150, 160, 161, 167, 169, 177, 185, and 198.
- 24. A recombinant endotoxin-neutralizing polypeptide according to claim 14, wherein the polypeptide contains the amino acid sequence of LBP
 25 having an amino acid substituted for an amino acid in a corresponding amino acid residue position of BPI.

- 92 -

- 25. A recombinant endotoxin-neutralizing polyp ptide according to claim 24, wh rein the amino acid substituted is at LBP amino acid residue positions 77, 86, 96, 118, 126, 147, 148, 158, 159, 161, 165, 167, 175, 183, or 196.
- 26. A recombinant endotoxin-neutralizing polypeptide according to claim 14, wherein the polypeptide contains the amino acid sequence of BPI having a cysteine residue substituted with an amino acid other than cysteine.
 - 27. A recombinant endotoxin-neutralizing polypeptide according to claim 26, wherein said cysteine residue is at BPI amino acid residue position 132, 135, or 175.
- 28. A recombinant endotoxin-neutralizing polypeptide according to claim 26, wherein the cysteine residues of BPI at positions 132, 135, and 175 are substituted with an amino acid other than cysteine.
- 29. A recombinant endotoxin-neutralizing
 20 polypeptide according to claim 14, wherein the polypeptide comprises a molecule which enhances the half-life of said polypeptide and is covalently bound to the polypeptide.
- 30. A recombinant endotoxin-neutralizing
 25 polypeptide according to claim 29, wherein said
 polypeptide contains a lipopolysaccharide-binding domain
 of BPI, LBP, a BPI variant, or an LBP variant.

- 93 -

- 31. A recombinant endotoxin-neutralizing polypeptide according to claim 29, wherein said molecule is an immunoglobulin fragment, a half-life enhancing portion of LBP, a half-life enhancing portion of an LBP variant, or polyethylene glycol.
- 32. A recombinant endotoxin-neutralizing polypeptide according to claim 29, wherein the endotoxin-neutralizing polypeptide of (a) is a C-terminal fragment of BPI and the molecule of (b) is an N-terminal fragment of LBP.
- 33. A recombinant endotoxin-neutralizing polypeptide according to claim 32, wherein said C-terminal fragment of BPI is a fragment having an amino acid sequence contained in BPI amino acid residues .15 60-456.
- 34. A recombinant endotoxin-neutralizing polypeptide according to claim 33, wherein said C-terminal fragment of BPI is BPI amino acid residues 60-456, 136-456, 277-456, 300-456, 200-456, 136-361, 20 136-275, 200-275, or 200-361.
 - 35. A recombinant endotoxin-neutralizing polypeptide according to claim 32, wherein said N-terminal fragment of LBP is a fragment having an amino acid sequence contained in LBP amino acid residues 1-175.
- 25 36. A recombinant endotoxin-neutralizing polypeptide according to claim 21, wherein said N-terminal fragment of LBP is LBP amino acid residues 1-59, 1-134, 1-164, 1-175, 1-274, 1-359, 1-134, or 1-197.

- 94 -

- 37. A recombinant endotoxin-neutralizing polypeptide of claim 18, wherein the p lypeptide further comprises a C-terminal fragment of LBP.
- 38. A recombinant endotoxin-neutralizing
 5 polypeptide of claim 23, wherein the C-terminal fragment
 of LBP is LBP amino acid residues 360-456 or 274-456.
 - 39. An isolated DNA molecule encoding a recombinant endotoxin binding polypeptide according to claim 14.
- 10 40. A vector comprising the DNA of claim 39.
 - 41. A transformed host cell comprising the DNA of claim 39.
- 42. A method for producing a recombinant endotoxin-neutralizing polypeptide according to claim 14,15 said method comprising the steps of:

culturing a transformed host cell comprising DNA encoding a recombinant endotoxin binding polypeptide according to claim 14, said DNA being operably linked to a promoter for expression of the polypeptide encoded by the DNA, said culturing being under conditions allowing expression of said polypeptide; and

isolating the recombinant endotoxin binding polypeptide produced.

- 43. A pharmaceutical composition comprising:
- a therapeutically effective amount of a recombinant endotoxin-neutralizing polypeptide according to claim 14; and
 - a pharmaceutically acceptable carrier.

- 95 -

- 44. A method of tr ating a subject suffering from an indotoxin-r lated disord r, said method comprising:
 administering to a subject having an endotoxin-related disorder a therapeutically effective

 5 amount of a recombinant endotoxin-neutralizing polypeptide according to claim 14, wherein LPS-mediated stimulation of neutrophils and mononuclear cells is inhibited.
- 45. A method of preventing an endotoxin-related disorder in a subject, said method comprising:

 administering to a subject a prophylactically effective amount of a recombinant endotoxin-neutralizing polypeptide according to claim 14, wherein the endotoxin-related disorder is prevented.

FIGURE 1A FIGURE 1C FIGURE 1D	
FIGURE 1A	
	l bi Schumany etal.)

2 = = 00 00 00 I.BP. : 1 (as reported by Schumany (correct sequence) zz < < L.BP-a L.BP-b Human Human Hamm

1/39

ΞΞ

လလ zz

FIGURE 1B

		<u> </u>	S		S	8 –	S	=	S	S	_	~	>	0	Ö	% ≃	≩	×	>	· œ	×	S	is.	<u>::</u> :	_ *	001
		-	S			-		<u> </u>	လ	ဟ	_	~	>	~			≩								¥	_
											9									7	20					
	L.HP-a	→	Ü	s:	ننـ	-	>	S	>	×	C	_	S	_	S		z			1		S	<i>≃</i>	S		
- Luniur	- -	C	Ü	s.	<u></u>	-	>	S	>		ŋ	_	S	_		>										
							<u>5</u>									<u> </u>	9									
		Ü	~	ے	۲	IJ	၁	>	- :	S	၁	S	S	٥	_	_ <	_	>		٥ /		Σ		_ _		
		Ü	~	_	;-	>	—	<	S	S	ပ	S	S	۵	_				<u>"</u>	- >			S (
						•	•	•	•																	
			150	0								_	9										0,			
limina	LBP-a	S	IJ	₹	-	_:	z	<u>-</u> :	<u>:-</u>	=	z	0	_	ш	S	×	<u>-</u>		>			<u> </u>	S	- ≃	_	
1	- BF-8	<u>:-</u>	Ü	≩	- :	- :	z		<u></u>	=	z	0	_					~ ~	×					~	Ü	• .
								9	_								-	90						75	25k	38 K
		n	Σ	-	0	×	S	>	S	တ	Ω	_		م	_ ≻	<u>ر</u>		_	۵,	>	+	:			_	_
		Ë	Σ	-	0	ᆇ	S	>	S		_	_:	0										=	-	_	_

FIGURE 1C

		*	20	õ		. 200							210	_								·	220	_			
Human	1 BF a	S.	<u>-</u>	<	_	-	=	> -	S	<u>-</u> :	>	<u> </u>	∢	<u>-</u>	~	<	÷	∢	9	Σ	<u>-</u> :	=	>	Ξ	<u>-</u> -	¥	
Humanı	÷	ν.	<u>-</u>	<	<u> </u>	-	=	>				=	<	<u>-</u>				∢	\sim		<u>.</u>	=	>	Σ	<u>:</u>		
								23(n								•	240	_								
		ŋ	Ħ	-	ഥ	Ξ	~	z	=		S	؎	>	⊢	_	_:	4	4	4		•		•	<u></u>	<u>=</u>	=	
		C	<u></u>	-	ت	=		Z	=	~					- :	_:	¥	4	>	Z	S	<u>-</u> :	<u>-</u>	<u>=</u>	<u></u>	=	
																			•		•	•	•				
			25	Q									×	_		•.							270				
Human	. H.	z	¥	χ Σ	>	>	<u>:-</u>	٧	-	S	_	>	>	ㄸ	Z	:	4	S	<u>-</u> `	>	>	=	=		C	>	
Himan	1.81. 6	z	¥	Z	>		: -	<		လ	<u>_</u>	>	>	Ŀ	z	: -	4	S		>	>	=	=	:::	C	>	
								28	0								•	290	_								
		<u>-</u>	z	<u>:</u>		- s	۲	_	=		-	ے	ے	<u>_</u>	S	z	_	~	_	<u>:-</u>	÷	×	<i>ۍ</i> :	<u>:</u>	~	_	
		-	Z			-	÷		<u>_</u>	Z	-	<u>-</u>		<u>_</u>	S		-	<u>-</u>			-	×	ν.	<u>.</u>	=	<u>-</u>	
			m Q	Q									310										320				
Homon	L.HP.:a	<u>:-</u>	>	_	~	VPRLARL	4	~	_	>	<u>م</u>	z	YPNMNL	z	ے	田口	ب	0	Ö	S	>	٥.	S	<	<u>-</u>	SAPL	
Human	4	<u>ت</u>	>	2	~	_	4	~	_	>	۵	Z	Σ	Z	_	Ľ	_	C	c	v		_	v	<	=	_	

Z Z

00

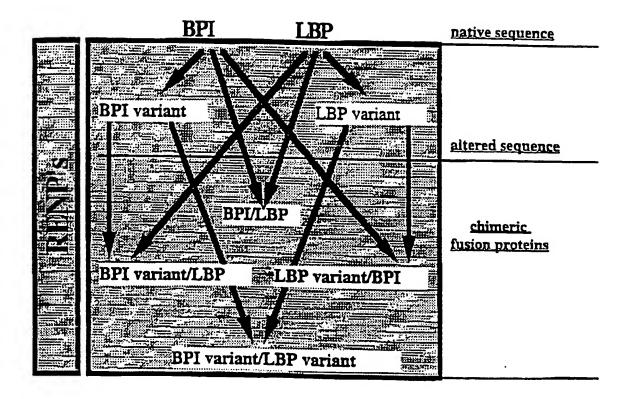
z z < <

Himan

FIGURE 1D

	٠,٠	٠,٠						_:		<u>.</u>	••				
				_	_		_	_			<u> </u>		_	_	
	· <i>y</i> r.			쏘			_	-			Ü			_	
	<u>-</u>		O	S			<	<	3		= :		<u>-</u>	<u>:-</u>	
	<u>-</u> :	_	370	:	÷		z	z	420	<	⋖		<u>-</u>	<u>-</u> :	
	<u>-</u> :	- :		z	z		<u>-</u>	<u>:-</u>		- :	-		÷	÷	
	>	> .		Ľ.	<u>:</u>		<u>.</u> :	<u>-</u> :		¥	×		=	=	
	<u>:-</u>	÷		<u>-</u>	:-		ij	ij		_	=		¥	¥	
_	<	<		_:	<u>-</u> :		>	>		z	z	2	=	=	
340	<u>_</u>	=		÷	÷	390	٠∠	¥		Ŀ	<u>:-</u>	440	_	-	
	-	-		<	<	1	S	s.		¥	×		~	C	
	<u></u>	<u></u>	•	S	S		<u></u>	<u></u>		ے	ے		<u>-</u> :	- :	
	Σ	Z		>	>		×	×		>	>		Ü	Ü	
	>	>	_		z		<u>-</u> :	<u>-</u> :		-	Ĺ.		-:	- :	
	<u>-</u>	<u>-</u>	360	H	:		<u></u>	<u></u>	014	H	÷		<u>_</u>	٥	
	a	=	·		∢		>	>	•		z		>	>	
	>	>		>	>		×	×			-		-	<u> -</u>	
	'n	S		S	S		>	>		_	-		ð	O	
_		- :		<u> </u>	_:	_		ㅗ		>-	>	^		>	
330	z	z		œ	×	380	င	IJ		> -	>-	430	~	~	
	Ü	C		ت	<u>:-</u>	•		<u>ـ</u> ـ		z	z		×	×	
	=	<u>-</u>		>	>		×	×			- :			-	
	S	S		=	_		<u> </u>	- :		<u> </u>	_;		<u>-</u> :	_`	
	<u>:-</u>	ä		<u></u>	<u></u>		<u>:-</u>	· ·			<		2	2	
	z	z	350	<u> </u>	'		Ü	U	4	<u> </u>	<u></u>		<u>~</u>	<u>-</u>	
	<u>-</u>	<u>-</u>		ۍ:	ော		÷	· :-		_	-		=	<u>-</u>	
				- III	- 4 - 1 - 1					L.H.	1.81%				
				11 miles	1					Human	Lumm				

Figure 2



ထမ 144 38 240 288 86 192 54 336 50 GTC GTC GAC AGC FIGURE 3A FIGURE 3C FIGURE 3C Het CTC Leu ACG TCA TAC AAC Aed GTG GTC 666 617 TAC ATG 66C 617 CAG GAC AGC AGA CTG CAG CCT CCT ATG Het TCC AAC AGC TCC AGG GTG GTC GCC AAG Lys 666 617 AGT FIGURE 3A TGG GCC TAC ATC Ile AAG Lys CCC CIC AGA GCG GAC AGG 666 61y E53 CAG CCG ACA CTG AAG Lys ren Len TGG SCG Ala GTG GGC CTG Leu CAT TTC TTT AAC Asd GAG GCC AAG Lys AAG AGG TGC ACC AAG Lys TTG CCT 715 TCC CAG AAG Lyb သည ATA ATC Ile CTG GAC TTT CAG AGG GCC AGG AGC GCT CDNA

97

BPI

145 39

533

289 87

38	13,	48(528 166	576 182	624 198	672 214	720	768 246
ATC Ile	AAT	CTC Leu	TGC Cys	GTC	CGA	TCC	ATA Ile	Acc
AAG P	GGC A	AAG C Lys L	AGC T	AAA G Lys v	CTT C	GTA T	AAA AY	GCA ACAIA TH
ATC A	AGC G	CTG A Leu L						
			TCC	AGC	GCG	TCT	Acc	CCA
AAT	ATG	GAT	TGC Cys	AAG Lys	TCT Ser	AAT	ATG	CCT
GCC Ala	AAA	GCT	ACC	TCA	GAG Glu	ACC	GTA	GCA
ASC	TY Lea	TCG	ATC Ile	ATC II8	ATT II0	GTC	CCA	GTG
AGC	TTC	ATT	Acc	CAC	AAA Lys	AAA Lys	CTC	E C
ATC Ile	AGA	TCC	CCC	GTG	AAA Lys	GAG	Act	GGT
TCC	AAG Lys	ATG Met	AAG Lys	CAC	CAC	TGC	CAG	TAT
TTC	CAA Gla	66C 61y	66c 61y	GTC	TTC	GTC	TTC	AAC
AAG Lys	GCA Ala	gyr gjn	TCA	AGT	CTC	CAG	TAT	ATC
rid Lid	AAG Lys	ATA Ile	Acg	AAC	CAA	AGC	CCT	GGA
66c 61y	TGG Trp	AGC	CCC	ATC Ile	ATC IIe	AAC	CAA	GCT
GTG Val	AAA Lys	CTC Leu	AAC	CAC His	CTC	ATG	CTG	GTG
AAT	GGG Gly	GAC	AGT	AGC	TGG Trp	AAG Lys	AAG	TCT
Pro	AGC	TTT Phe	660 61 y	AGC	666 61y	AAC	TCC	GAT
103	385 119	433 135	481 151	529 167	577 183	625 199	673 215	721

FIGURE 3B

FIGURE 3C

816 262	864 278	912	960 310	1008 326	1056	1104	1152	1200	1248
GAG	CCC	TTC	ACC	ACC	AAC	TCT Ser	GCC	66C 61y	Cert Leu
AGT	TTT	TTC	ATG	ACA	CCC	CTG	CAG	ATT	AGG
TAC	GAG	TAC	AAG Lys	CTG Leu	TTT	CAC His	GTC	CTG	AAC
TTT	ATG	GAC	TTG	CGA	AAG Lys	CCA	GAT	TTC	TCC
GAG	GTG	TCA	GTC	TTT	AAG Lys	CCG	GTG	C T T	GAG
666 61y	Pro	CTC	666 61y	AAA Lyg	GCC	ACC	GCC	TCC	GCC
AAG	CCA	GGC	GCT	TCC	GTG	TCC	CCT	GCT	AGC
ATG	GCT	7.5 2.5 2.0	GAG	GAG Glu	GAG	GCC	TAC	CTC	GTC
CAG	TTT	TAC	CAA	AAG Lys	CCT	TCA	TTC Phe	TCC	GAG
GTA	CCC Pro	GTA	TAC	CCA	CTA	GTC	ACC	TCC	ATG
GAT	Pro	ATG	GTA	ATT	TTC	CAT	rea Lea	AAC Asn	TCC
Sec.	CCA	CGC	C. S.	ATG Het	Acc Thr	ATC Ile	GGC	CCC	GGT
Acc	AAT	GAC	666 61y	GAC	GGA G1y	CAG	ACC	CTC	ACT
GAG	CAC	CAT	GCC	GAT	TTT	ATA Ile	CCC	GTC	ACA
GCT	CAC H1s	GCC	ACA Thr	AGA	TTC Phe	AAG Lys	CAG	GCC	CAC
ACG	AAC	GCT	AAC	Car	AAG Lys	ATG	GTG	Crr Len	ATG
769	817	865 279	913	961	1009	1057	1105	1153	1201

FIGURE 3D

1296	1344	1392	1440	1488 486	1536 488	1584	1632	1680	1728	1776	1813
TCA	TAC	AAA	GTG	TAT	ככד	CAG	YYY	GTG	ATA	TTG	
CAC H18	AAC Asn.	CAG Gln	GTA	GTC	GTT	CTC	CAG	GGT	GAG	CAT	
AAG Lys	ATG	Cty Len	AAC	GTT	CCT	CCT	ATT	CAT	CCA	TTC	
CTC Leu	ATC Ile	AAA Lys	TAC	GAC	GCA	AAT	CAG	GTG	GCT	MAT	U
GAA	GAT	GAG	CTC	GCA	၁၁၅	999	ACT	*	MAG	AGA	TGT
CTC Lea	CAG	AAC	CAG Gln	GGT	JJ.	Ç	TCG	GGA	ट्टिं	CCA	TCA
CTC Leu	23	GTT	GTC	TTC	23	TCC	TCT	ATT	AGG	TX	TET
7.50 J	13	AGG	AGA	Sign I	99	E	Act	TTC	TCA	TTG	TIT
AGG	GAA	CCC	GCC	7. 2. 2. 2.	990	TGC	3	TTA	E C	3	TGG
GAT	GTT Val	CTG Leu	CCG	Phe	TGC	၁၅၅	TTG	ACA	r T	TIT	TTC
222	CCG	GTG	ACG	AAC	999	ACC	၁၁၁	CAA	GAG	GTG	MC
AAG Lys	TTC	53	CCG	CAG Gln	CAG	၁၅၅	AGC	GAG	TAT	ATC	3
Cro Lea	CCC	ATT	Seu Seu	CAC	CAC	TGG	MG	CAC	GAT	GGA	*
GAG	66C 61y	CCC	CCT	CCT	AGG	CTG	ACC	¥	AGG	CCA	ATG
GGA Gly	ATT Ile	GTA	TTC Phe	CAG Gln	TGA	999	TTA	TCT	TIT	CC	TTC
GTT	AAT	ATT Ile	GGC Gly	E a	AAA Lys	GAT	ATC	TGA	TAT	TIT	TGC
1249	1297	1345	1393 455	1441	1489	1537	1585	1633	1681	1729	1777

							u.	FIGURE	3E 4A				FIGURE	4 A B			
ב	LBP	Ехр		ression	clone	Q							GURE	4			
	CCT A	CCT ACC Nhel	CCA	्र ट्रा	CAC	166	SA S	TCT	AGG	ATG	666 61y	GCC Ala	TTG	GCC	AGA	GCC	48
2 8	CTG Leu	CCG	TCC	ATA Ile	77. Leu	755 250 250	GCA	TTG	CTG Leu	CT.	Acg	TCC Ser	ACC	CCA	GAG Glu	GCT	96
97	CTG	GGT Gly	GCC	AAC	CCC	66c	233	GTC	GCC	AGG	ATC	ACC	GAC	AAG Lys	GGA G1y	CTC	144
2 4 0	CAG	TAT	GCG	GCC	CAG	GAG	666 61 y	ray Lea	110	GCT	Z Z	CAG	AGT	GAG Glu	CTG	CTC	192
56	AGG	ATC	ACG	ក្តក្នុ	CCT	GAC	TTC	ACC	666 617	GAC	13	AGG	ATC Ile	CCC	CAC	GTC	240
41	66C 61y	CGT	666 61y	CGC	TAT	GAG	TTC	CAC	AGC	CTG	AAC	ATC Ile	CAC	AGC	TGT	GAG Glu	288
6 8	CTG	5.5	CAC	TCT	GCG	15 de	AGG	CCT	GTC	CCI	66c 61y	CAG	66c 61y	CTG Leu	AGT	CTC	336 103
17	AGC	ATC Ile	TCC	GAC	TCC	TCC	ATC Ile	CGG	GTC	CAG Gln	66C 61Y	AGG	TGG Trp	AAG Lys	GTG	CGC	384 119
202	AAG	TCA	TTC	TTC	AAA	CTA	CAG	66C 61y	TCC	TTT	GAT	GTC	AGT	GTC	AAG Lys	66C 61y	432
6 2	ATC Ile	AGC	ATT IIe	TCG	GTC	AAC	23	24 25 26 26	77G	66c 61y	AGC	GAG	TCC	TCC	666 61y	AGG	480

FIGURE 4B

528 167	576 183	624 199	672 215	720	768 247	816 263	864	912 295	960	1008
GAG	AAC	GAA	ACT	AGC	TTT	CTT	TTT	GAG	GAC Asp.	CGG
GTG	CAC	TGC	CAA	TAT	ATG	CTC	TAC	CAT	CCT	CCA
GAC	TTC	ATT Ile	CTC	GAT	GTG Val	ACC	GTC	TAT	CCG	GTC
GCT	252	AGG	TAT	ATT	GAG	GTT	ATG	GTT	ATA Ile	TTC
ATC I10	AAC	AGC	CCT	GAC	77.5 15 c	CCCA	AAA	Cord Leu	ATG	CCC
GAC	13	GAG	CAG	GCC	ATG	TCT	AAC	AGC	GAC	CGA
AGT	CTC Draga	273	Eg Eg	TTC	CAG Gln	CGT	CAC	GCC	GAT	TTC
AGC	166 1rp	GTA	GAT	AGT	GCC	CAC	GAA	Acg	ACA	TCC
760 Cys	666 61y	MAA	TCC	GAC	ACA	AAC	GAG	AAC	ATC Ile	AAG Lys
AGC	TIC	CAG	TCC	ATT	GCA	CGT	CCT Pro	TTC	TCC	Acc
Ser	GAQ Asp	TTC	GTG	GAG Glu	CGG	CAT	Len	GTC	TTC	Acc
SSS	GGA	AAG Lys	TCG	ACA	CCT	TTT	AGC	TAT	AAC	CTC
E E	TCG	TCC	AAA Lys	ACA	GCC	ATC	ATG	GAT	CTG	CGA
GII Kel	ATG Het	GAG	CAG	GTT	GAA Glu	GAA	GIC Val	TCG	TAT	ATC Ile
ACA	GAC	ATT Ile	ATC Ile	CCA	GTG	GGT	GCA	ATC Ile	GGA G1y	AAT
CCC	GTG	CAG	ATG	crc	TTA	AAG Lyb	GCT	GCC	GAA	TCT
181	529 168	577	625	673 216	721	769 248	917	865 280	913	961

7 * 7	1104	1152 375	1200	1248	1296 423	1344	1392	1440	1488 482	1500
t J	GAC	AAG Lys	TTG	GTA	CTG	AAG Lys	CGT	TTC	TGA	
• •	GTG	AGC	ACC	AAG Lys	GAG	CCC	AAG	CTG Leu	AGA	
;	TCT Ser	TCC	GCC	GGA	GCA	TAC	CTG	TTC	CAA	
3	CTC	AGC	TCC	CCA	AAT	TTC Phe	CTG	GAC	CAA	
2	AAT	CCC	GTG	AAG Lys	TTC	Acc	CCT	AAG	GGA	
3	666 61y	252	AAT	CTG	E G	AAC	£3	CAT	TGA	
	CCT	Sec.	Act	TTC Phe	GGA	53	CCC	ATC Ile	GTT	
ABD	AGC	GTG	GCC	666 61y	GTT	ATC	TTC	CAG	AGA	
Met	TTC	TTT	GTG	ACT	MAA	TAC	GGC	CTC	ATG	
ASH	AAC	GCC	AGT	ATC Ile	TCC	TAT	GAA	666 61y	TAC	
Pro	CTG Leu	GAT	CTC Lea	AAG	GYA	AAC	GCC	region 1	GIN	
Tyr	Ser Leu	ATA	CGG	AGC	AAA Lys	CTC Lea	TTG Leu	GAC	GTC	•
Ze Z	CCG	GAG	TTC	Acc	CTG	CTC Leu	AAG Lys	TAC	AAT	TC GAG XhoI
Arg	GCT	ATG	GTC	AAT	GYA Glu	GCG	GAT	CTC	GCC	STS SPO
Ala	TCT	TAT Tyr	CCT	TTC	GTG	GAA	AAT	CAG	GGT	TTG (
Len	CCC	CCC	GAG	ACC	AAA Lys	77.5 Leu	TTC	GTT	TTG	AGC
328	1057	1105	1153	1201	1249	1297	1345	1393	1441	1489

		Ξ	Ξ	: -	==	-		-		=3	ະວ	=0	=			•	_:		`	_:	_:	•
		•		Ξ	=	-				=41	_	_	::	_			∢	<	V	<	۰	
		•	•	=	:			-		_	==	==	_	_			>	<u>-</u> :	<u>.</u> :	4	_;	
		•	•	ย						3	3	ย	>	>			<u> -</u> :	_:	_;	۲	>	
-30		•	•	-				_		=	<u>-</u>	=	=	=		•	၁	C	ဗ	ŋ	5	•
		. '	•	E							e	_	-	_	 20		ш	<u>ت</u>	m	0	0	_
										v.	=	y.	20	E			ᆇ	~	0	0	0	_
										_	=	_		_			∢	V	4	S	ပ	
SINISION SINISII SINISION SINI								-		_		_	=	=		•	¥	∢	⋖	4	4	•
URE URE URE							9	: -	••	_	_	_	>	G		•	>	>	>	>	>	•
								•	•	-	-	_	_	_			V	ш	0	۵	۵	_
										23	v:	æ	>	>		•	_	_	_	_	ے	•
								-	••	_	-	_	Ξ	>		*	Ö	ט	Ö	Ö	Ö	*
5A								•	•	-	_		_	-		•	¥	¥	×	×	¥	•
FIGURE										-	-		S	-		-	۵	۵	۵	0	0	_
OG O								-	••	S	s	S	>	G	2		⊢	—	⊢	S	-	
正										٩	8	ے	3	3		•	-	_	-	_	-	•
								-	-	-	-	-	-	_		•	~	~	~	~	~	•
							_				-	•	Ф	-			4	-	4	>	4	
							.20	-	••		a	G	G	G			>	-	>	>	>	
											-	_	=	-			>		_1	>		
								_	••		G	G	ပ	7		*	Ö	Ö	G	Ö	Ö	•
											3	_	C	<u>۔</u>		•	_	<u>-</u>		۵.	_	•
											_		ಬ	50		•		z				*
											60	20	_	_	 _		>	—	⋖	>	_	
	-87	1.187	isi	H	<u> </u>												J.B.	1.130	- E	Tag	131.1	
	Monse	Rubbit	Human H	floor:	Bovine												Monse	Rabbit	Heman	Human	Bovine	

FIGURE 5B

14/39

```
0000.8-
3
   ~~~~~
                    0 C > x
  CCZA
  -- < = = = >
              2 --
                   4 4 2 4
                                  \times \times \times \times \times
                   K K S K
                                  ~ ~ 0 ~
                   F 4 0 0 --
   -----
                  S S S S --
                                  3 3 3 3 *
2 - - - - - s z
                 D Z I S Z
                            90
  - 5 5 6 6 6
                -- 4 -- 1 -- 4 --
                -- C C C C Z
                 ひにひにに!
   2 2 2 2 2 9
                 Z M N E C
                                  < x < < --
                  □ × × □ >
   8 -- > -- 2 × =
                • S S S S • 8
 • ------
   医医肠链链性
                • " " " " " " •
   ~ × × × × --
 • 22224.
```

Mouse LBP Rabbit LBP Human LBP Human BPI Bovine BPI

```
.×≃ ♡ ×
                                                                                                                                                                                                                                                                                                                                                                                                                                                                                               -- o o o ≃
                                      • 000 • 5
                                                                                                                                                                                                                                                                                  120 T T T W T -
                                                                                                                                                                                                                                                                                                               Z C S F
                                                                    -- > > < < -- <del>1</del> -- -- <del>1</del> --
                                                                                                                                                                                                                                                                                                             0 < Z Z --
                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                 FIGURE 5C
                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                       4 S S S --
                                                                                                                                                                                                                                                                                                                                                                                                                                                         -00××
                                                                                                                                                                                                                                                                               -- Z Z O O --
                                                                   • 0 0 0 0 • 130
                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                               \Rightarrow \geqslant \geqslant -
                                                                                              בטטט --
                                                                                             G S Q K
                                                                                                                                                                                                                                                                             • 0000.
                                                                                           1.8P
1.8P
8P1
8P1
                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                 Rabbit
Human
Human
Bovine
                                                                                                                                                                                                                                                                                                                                                                                                                                                                                                              Rabbit
Unman
Numan
Bovine
```

```
- > > > < 00 -- -- -- --
                 • 5555.
 - Z Z O -
                    v ⊞ 4 4 --
 -· > > > >
                    以目目以
  - X Z F Z - Z Z Z Z
            < > > <
  2 Q 11 Z
 - v v O O -
        240 · - - × × -
 • zzzz •
  • # # # # #
                 260
50
50
50
50
50
                    • ~ ~ ~ ~ ~ •
 - s s o s · = = = - .
 · > > > > · A Z Z 4
                   0 4 0 0 –
 -- a c z a
       230
          - = × × --
 ---->
                   ××××- - Z Z × Z
                   ν z = = --
  Rabbit
Numan
Human
Kovine
                     Rabbit
Buman
Human
Bovine
```

```
- x x x x -
      ÷ ∨ ∨ ≯
 - C C > - · < < < < ·
     B 0 0 0 -
 -- 00==
E C S
 Z <del>Z</del> <del>Z</del> <del>Z</del> <del>Z</del> ... - Z Z Ω ω
K Q S S -- Q E R L
      -- -- -- > 5
            - 22 4 4
 Rubbit
Burnan
Brocke
             Rabbit
Bunan
Bunan
Bovine
```

FIGURE 5F

```
Rabbit
Human
Duman
Ravine
                                              Rabbit
Henran
Henran
Horine
```

FIGURE 6

								ပု
09	QSELLRITLP 120	RVQGRWKVRK	180 SGDLGWLLNL	25/30 230 240 EIDSVAGINY GLVAPPATTA	300 LVYQEAGVLK	360 VQPTGLTFYP	420 LKHSNIGPFP	479 LFGADVVYK*
50	YAAQEGLLAL 110	ISSUSISISI		25/30 230 EIDSVAGINY	290 LSDYFFNTAG	350 VSASTPPHLS	410 ELKLDRLLLE	470 VVLQPHQNFL
40	VARITDKGLQ 100	SALRPVPGQG	160 RPTVTASSCS	220 PYLQTLPVTT	280 AAHDRMVYLG	340 KFPNMKIQIH	400 VSAESNRLVG	460 PTPARVQLYN
0	20	-						_
3	PEALGANPGL 90	LNIHSCELLE	150 NLLLGSESSG	210 IQKSVSSDLQ	270 PFAPPVMEFP	330 FGTFLPEVAK	390 IGMHTTGSME	450 NEKLQKGFPL
20 3	ILLALLITET PERLGANPGL VARITDKGLQ YAAQEGLLAL QSELLRITLP 80 90 100 110	LNIHSC	140 VSVKGISISV NLLLGSESSG	IQKSVSS	260 270 280 290 300 FYSENHHNPP PFAPPVMEFP AAHDRMVYLG LSDYFFNTAG LVYQEAGVLK	520 330 340 340 350 360 360 SKFRLTTKF FGTFLPEVAK KFPNMKIQIH VSASTPPHLS VQPTGLTFYP	180 180 410 420 ANSSLASLFL IGMHTTGSME VSAESNRLVG ELKLDRLLLE LKHSNIGPFP	1VPILVLPRV NEKLQKGFPL PTPARVQLYN VVLQPHQNFL LFGADVVYK* -C
	N- <u>MGALARALPB ILLALLITBI PEALG</u> ANPGI 70 80 90	DETGOLRIPH VGRGRYEFHS LNIHSCELLE	NLLLGS					

FIGURE 7

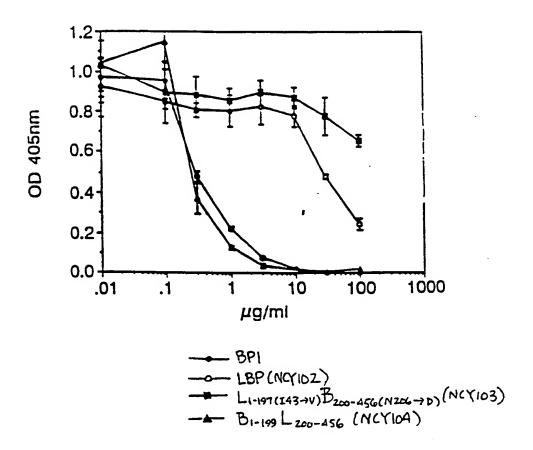
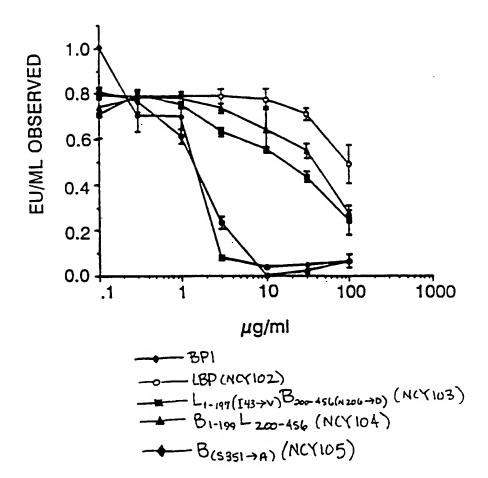


FIGURE 8



WO 96/34873 PCT/US96/06134

22/39

FIGURE 9

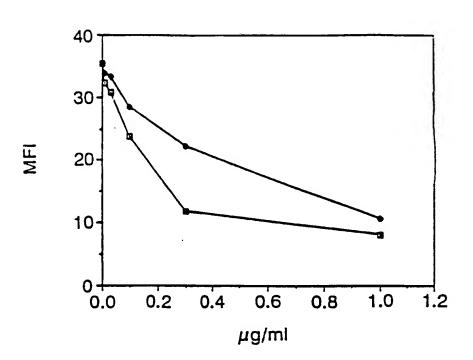
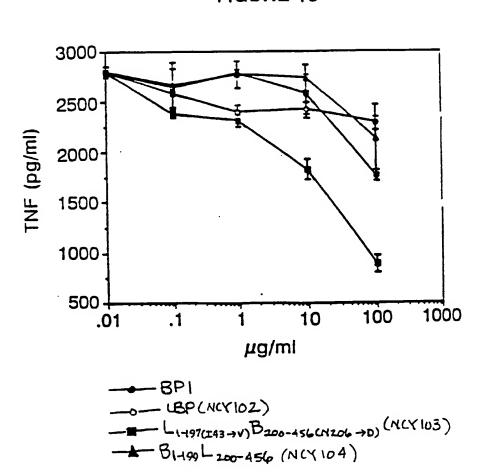
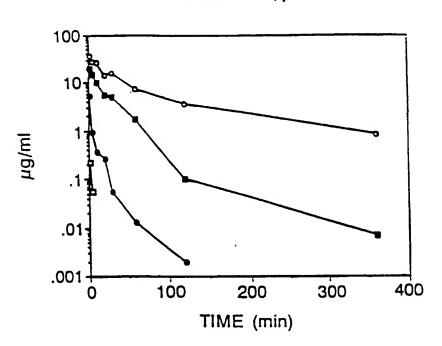


FIGURE 10

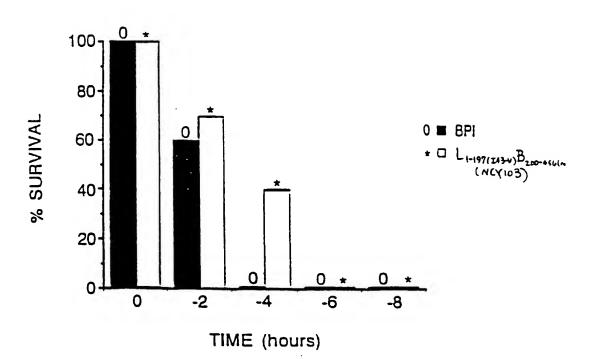






WO 96/34873

FIGURE 12



WO 96/34873

FIGURE 13A

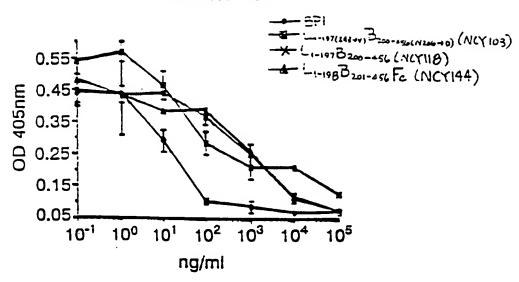


FIGURE 13B

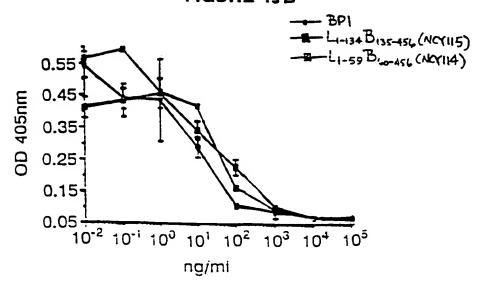
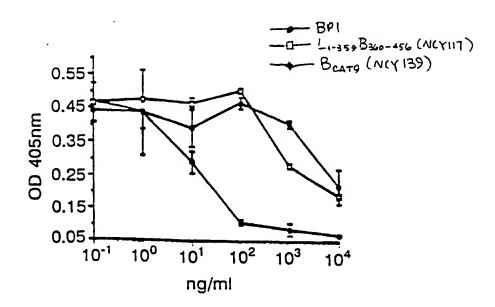
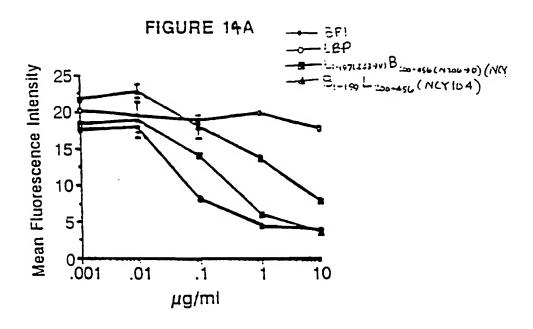


FIGURE 13C







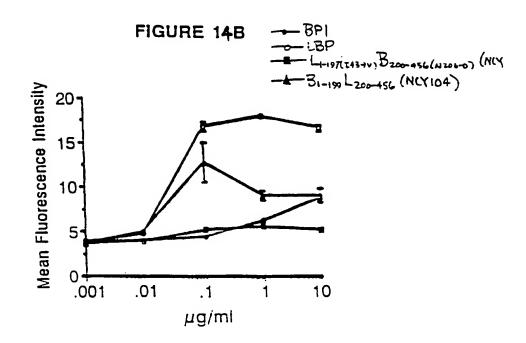


FIGURE 15

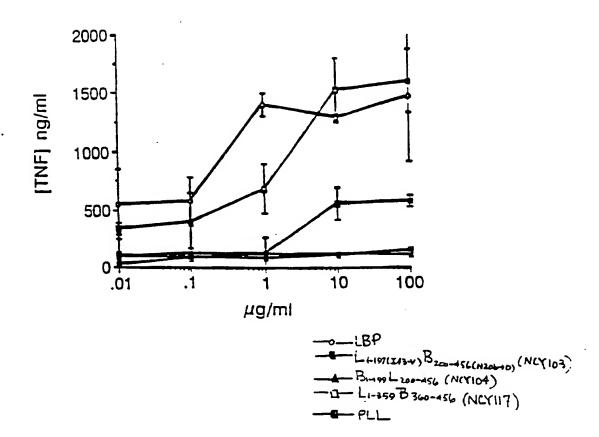
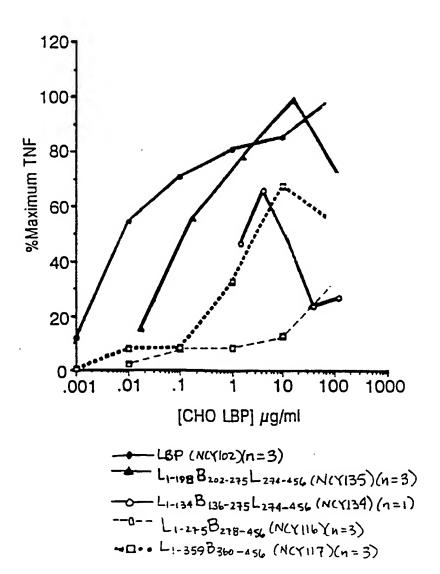


FIGURE 16



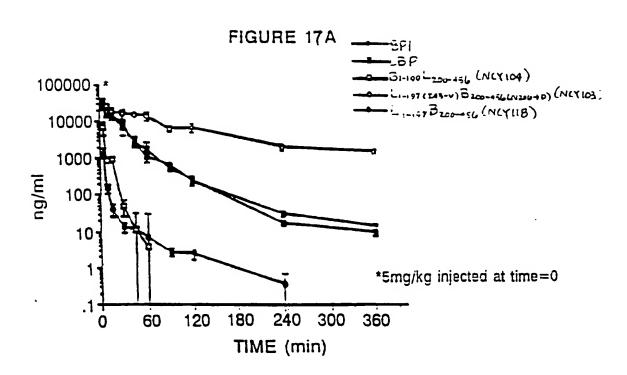


FIGURE 17B

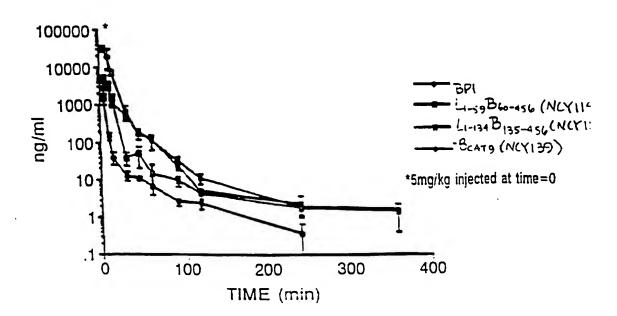


FIGURE 17C

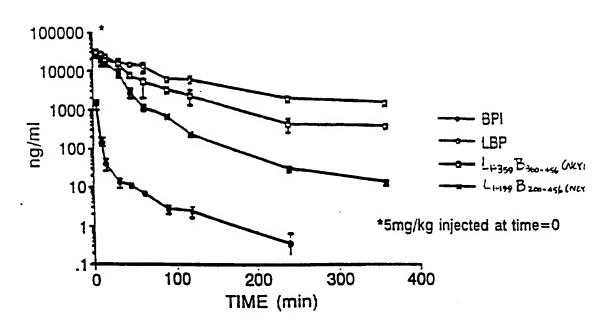


FIGURE 17D

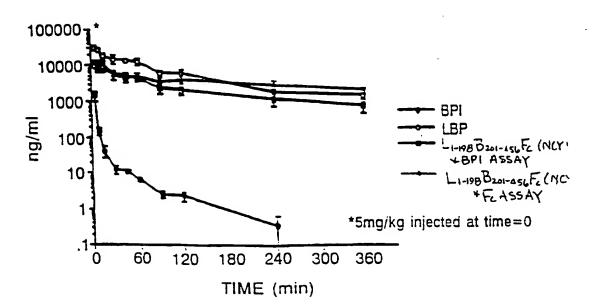


FIGURE 17E

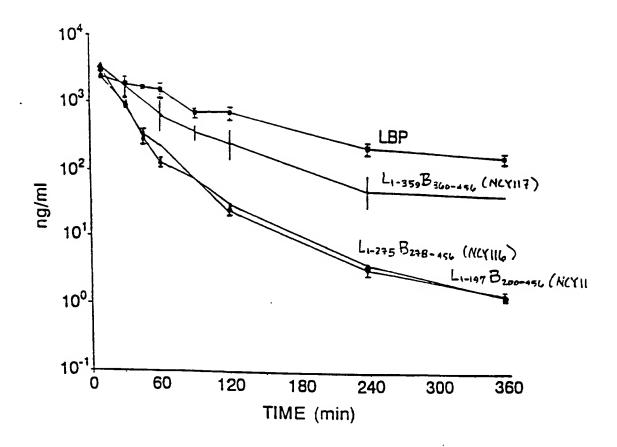


FIGURE 17F

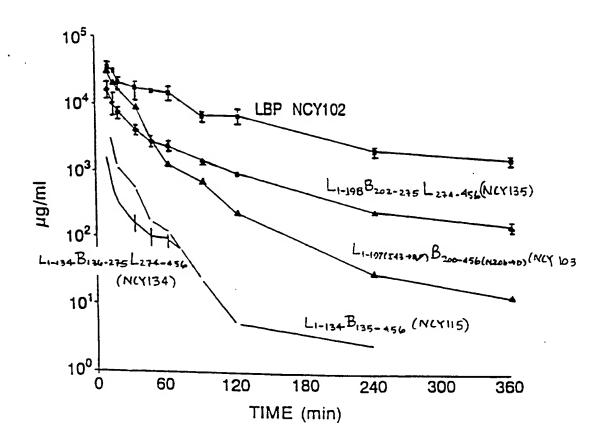
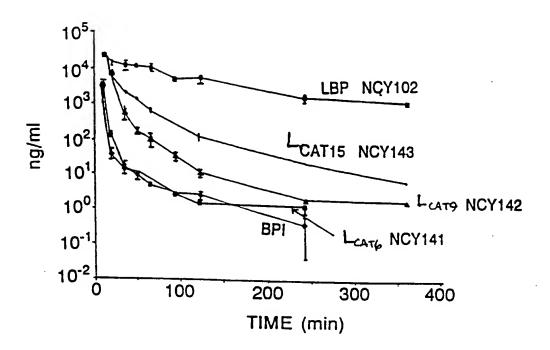


FIGURE 17G



5mg/kg compound injected i.v. at t=0

FIGURE 17H

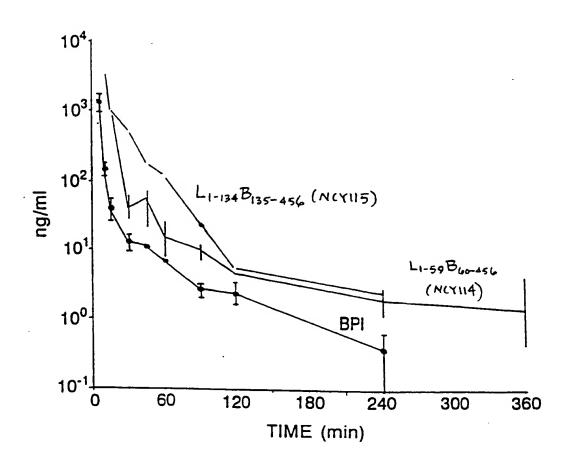


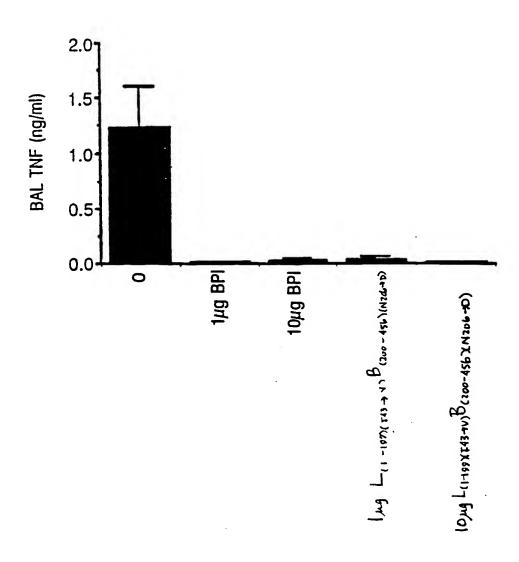
FIGURE 18

Lane Number

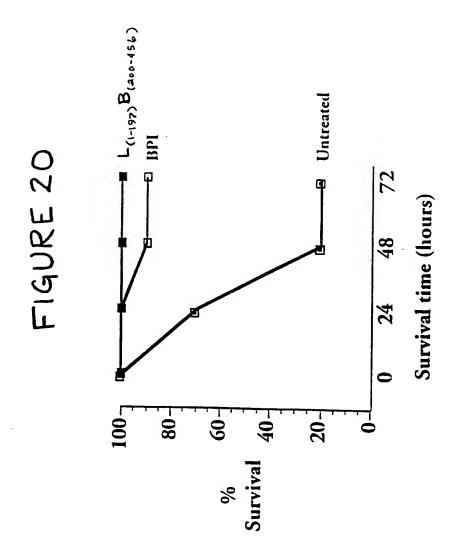
1 2 3 4 5 6 7 8 9 10 11 12



FIGURE 19



.



INTERNATIONAL SEARCH REPORT

International application No. PCT/US96/06134

		- <u></u>							
	ASSIFICATION OF SUBJECT MATTER								
IPC(6) US CL	:C07H 15/12; C12P 21/06; A61K 39/00, 38/00; :530/300, 350; 435/69.1, 320.1, 252.3; 536/22.1	C07K 1/00							
	to International Patent Classification (IPC) or to bo	, 22.3; 314/2 oth national classification and IPC							
	LDS SEARCHED								
Minimum	documentation searched (classification system follow	ved by classification symbols)							
	530/300, 350; 435/69.1, 320.1, 252.3; 536/22.1,								
		22.3, 314/2							
Documenta	tion searched other than minimum documentation to	he extent that such documents are included	in the fields searched						
			in the netus scarence						
Electronic	data base consulted during the international search (name of data base and, where practicable	, search terms used)						
DIALOG			, search terms used,						
2 - 20									
	CUMENTS CONSIDERED TO BE RELEVANT								
Category*	Citation of document, with indication, where	appropriate, of the relevant passages	Relevant to claim No.						
X	WO,94/25476 A (SCOTT et al.	10 November 1994, see	1 20 42						
	entire document.	10 140 verilber 1994, see	1-38, 42						
X	WO 94/18323 A (THEOFAN et	al) 18 August 1994, see	14, 31, 32,						
	entire document.	, 1, 1, 1, 1, 1, 1, 1, 1, 1, 1, 1, 1, 1,	11, 01, 02,						
· .			·						
×	US 5,348,942 A (LITTLE et al) 20	September 1994, column	14, 31, 32, 33						
	5, lines 40-45, column 7, lines 3-	23.	,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,						
-									
Ī									
1									
l									
Furth	er documents are listed in the continuation of Box (See patent family annex.							
	cial categories of cited documents:	"T" later document published after the inter	national filing date or priority						
"A" doc to b	ument defining the general state of the art which is not considered e of particular relevance	date and not in conflict with the applicat principle or theory underlying the inve	tion but cited to understand the						
	ier document published on or after the international filing date	"X" document of particular relevance: the	claimed invention cannot be						
'L' doc	ument which may throw doubts on priority claim(a) or which is	considered novel or cannot be consider when the document is taken alone	ed to involve an inventive step						
cito	to establish the publication date of another citation or other citation (as specified)	"Y" document of particular relevance; the	claimed invention cannot be						
O° docu	ument referring to an oral disclosure, use, exhibition or other	considered to involve an inventive combined with one or more other such	step when the document is documents, such combination						
	ns ument published prior to the international filing date but later than	being obvious to a person skilled in the art							
the	priority date claumed	'&' document member of the same patent f							
Date of the a	ctual completion of the international search	Date of mailing of the international sear	ch report						
08 JUNE 1	996	16 JUL 1996							
Vame and ma	ailing address of the ISA/US	Authorized officer							
	er of Patents and Trademarks	Authorized officer C Ts ~ fir							
Washington,	D.C. 20231	H. F. SIDBERRY							
Facsimile No	(703) 305-3230	Talaska No. (703) 000 010:							

τ

INTERNATIONAL SEARCH REPORT

International application No. PCT/US96/06134

Box I Observations where certain claims were found unsearchable (Continuation of item 1 of first sheet)
This international report has not been established in respect of certain claims under Article 17(2)(a) for the following reasons:
1. Claims Nos.: because they relate to subject matter not required to be searched by this Authority, namely:
2. Claims Nos.: because they relate to parts of the international application that do not comply with the prescribed requirements to such an extent that no meaningful international search can be carried out, specifically:
3. Claims Nos.: because they are dependent claims and are not drafted in accordance with the second and third sentences of Rule 6.4(a).
Box II Observations where unity of invention is lacking (Continuation of item 2 of first sheet)
This International Searching Authority found multiple inventions in this international application, as follows:
Please See Extra Sheet.
1. As all required additional search fees were timely paid by the applicant, this international search report covers all searchable claims.
2. As all searchable claims could be searched without effort justifying an additional fee, this Authority did not invite payment of any additional fee.
3. X As only some of the required additional search fees were timely paid by the applicant, this international search report covers only those claims for which fees were paid, specifically claims Nos.: 14-38, 42
4. No required additional search fees were timely paid by the applicant. Consequently, this international search report is restricted to the invention first mentioned in the claims; it is covered by claims Nos.:
Remark on Protest The additional search fees were accompanied by the applicant's protest. No protest accompanied the payment of additional search fees.
- F-Julian of additional section (ces.

INTERNATIONAL SEARCH REPORT

International application No. PCT/US96/06134

BOX II. OBSERVATIONS WHERE UNITY OF INVENTION WAS LACKING This ISA found multiple inventions as follows:

This application contains the following inventions or groups of inventions which are not so linked as to form a single inventive concept under PCT Rule 13.1. In order for all inventions to be examined, the appropriate additional examination fees must be paid.

Group I, claim(s) 1-4, 9-13, 43, drawn to method of in vivo detection of a site of Gram-negative bacterial infection, labelled endotoxin neutralizing polypeptide, composition, classified in Classes 514, subclass 2.

Group II, claim(s) 5-8, drawn to a method of in vitro detection of a Gram-negative bacterial infection, classified in Class 436, subclass 517.

Group III, claim(s) 14-38, 42 drawn to endotoxin-neutralizing polypeptides, method of making the polypeptides classified in Classes 530 and 435, subclasses 300, 350; 69.1.

Group IV, claims 39-41, drawn to DNA encoding a recombinant endotoxin binding polypeptide, vector and host cells, classified in Classes 536 and 435, subclasses 22.1, 23.1, 23.5 and 320.1 and 69.1.

Group V, claims 44, 45, drawn to methods of treatment using an endotoxin neutralizing polypeptide, classified in Class 514, subclass 2.

The inventions listed as Groups I, II, III, IV, IV do not relate to a single inventive concept under PCT Rule 13.1 because, under PCT Rule 13.2, they lack the same or corresponding special technical features for the following reasons: Group I is directed to a labelled endotoxin neutralizing polypeptide which is not defined. The polypeptide may be a variant of BPI or LPS in that the amino acid must not be identical to the amino acid sequences of either BPI or LPS. The methods of Group I and II are in vitro or in vivo, the method steps will differ in parameter, reagents and method steps.

The invertions of Group III are directed to polypeptides which are different from that required in Groups I and II to preform the methods of detection. The polypeptide of Group III is not the special technical feature set forth in the inventions of Group I and II. Group IV is directed to DNA which encodes a an endotoxin neutralizing polypeptide. However, variants of BPI and LPS occur spontaneously or may be achieved by chemical modifications. Group V is directed to other methods which use the endotoxin neutralizing polypeptide, but not one which is labeled as of Groups I and II.

The special technical feature which links the inventions is known in the art, for Groups I, II and III, as SCOTT et al WO 94-25476, discloses an endotoxin neutralizing polypeptide wherein the serine at 351 has been changed to alanine. (see page 18, lines 22-27. Other variants of BPI are disclosed by SCOTT et al at page 35.

This application contains claims directed to more than one species of the generic invention. These species are deemed to lack Unity of Invention because they are not so linked as to form a single inventive concept under PCT Rule 13.1. In order for more than one species to be examined, the appropriate additional examination fees must be paid. The species are as follows:

(a)species of BPI variants which are chimeric: claims 29-35 (b)species of BPI variants which have substitutions: 14-28, 36-38.

The species listed above do not relate to a single inventive concept under PCT Rule 13.1 because, under PCT Rule 13.2, the species lack the same or corresponding special technical features for the following reasons: the first species are directed to constructs comprising BPI or LPS and/or an immunoglobulin fragment. The second species is directed to BPI variants which have substitutions in the amino acid sequence. The variants have only substitutions. The chimeras are comprised of the LPS or the immunoglobulin fragment and thus are comprise an element not need in the variant BPI proteins.

•

D

ŧ